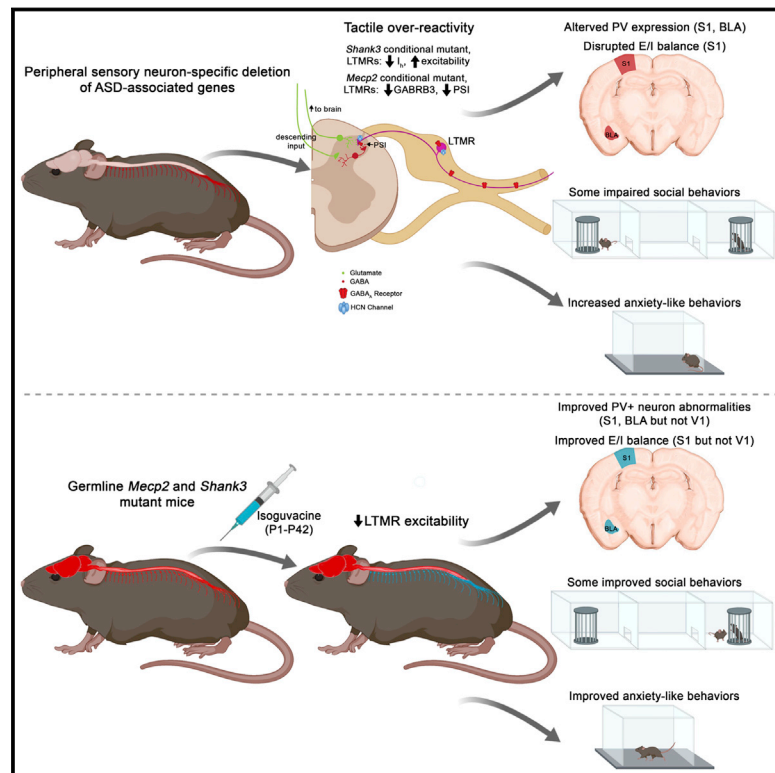


Targeting Peripheral Somatosensory Neurons to Improve Tactile-Related Phenotypes in ASD Models

Graphical Abstract



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In Brief

Treatment with a peripherally restricted GABA_A receptor agonist in multiple distinct autism spectrum disorder mouse models reveals a potential therapeutic strategy for select ASD-related behaviors.

Highlights

- Distinct pathophysiologies cause peripheral sensory neuron dysfunction in ASD models
- Peripheral somatosensory neuron dysfunction alters brain development and behavior
- A peripherally restricted GABA_AR agonist, isoguvacine, reduces tactile sensitivity
- Chronic isoguvacine treatment improves a subset of ASD-related phenotypes in mice



Targeting Peripheral Somatosensory Neurons to Improve Tactile-Related Phenotypes in ASD Models

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SUMMARY

Somatosensory over-reactivity is common among patients with autism spectrum disorders (ASDs) and is hypothesized to contribute to core ASD behaviors. However, effective treatments for sensory over-reactivity and ASDs are lacking. We found distinct somatosensory neuron pathophysiological mechanisms underlie tactile abnormalities in different ASD mouse models and contribute to some ASD-related behaviors. Developmental loss of ASD-associated genes *Shank3* or *Mecp2* in peripheral mechanosensory neurons leads to region-specific brain abnormalities, revealing links between developmental somatosensory over-reactivity and the genesis of aberrant behaviors. Moreover, acute treatment with a peripherally restricted GABA_A receptor agonist that acts directly on mechanosensory neurons reduced tactile over-reactivity in six distinct ASD models. Chronic treatment of *Mecp2* and *Shank3* mutant mice improved body condition, some brain abnormalities, anxiety-like behaviors, and some social impairments but not memory impairments, motor deficits, or over-grooming. Our findings reveal a potential therapeutic strategy targeting peripheral mechanosensory neurons to treat tactile over-reactivity and select ASD-related behaviors.

INTRODUCTION

Autism spectrum disorders (ASDs) are a heterogeneous group of complex neurodevelopmental disorders characterized by impairments in social communication and interactions and restricted and repetitive behaviors. Although rates of ASD diag-

nosis are increasing, with approximately 1 in 59 people in the United States reported to be living with ASD, there are no Food and Drug Administration (FDA)-approved treatments for core ASD symptoms (Baio et al., 2018). Furthermore, a majority of adults with ASD (60%) exhibit concerns about current medication options aimed at alleviating co-morbid ASD symptoms due to a lack of efficacy and adverse side effects, including fatigue and sedation (Howes et al., 2018). Of note, pharmacological modulators of GABA receptor signaling have gained attention for possible therapeutic utility in patients with ASD. Bumetanide (a selective chloride importer NKCC1 antagonist) and arbaclofen (a GABA_B receptor agonist) are being explored as means of restoring GABAergic inhibition and may show promise in recent clinical trials with regards to improving social behaviors in children with ASD, although side effects of fatigue and irritability pose concerns (Erickson et al., 2014; Hadjikhani et al., 2018; James et al., 2019; Lemonnier et al., 2017; Veenstra-VanderWeele et al., 2017). New therapeutic strategies to treat symptoms observed in ASD patients and reduce off-target effects are clearly needed.

A major hurdle in developing effective treatments for ASD patients is that symptoms and severity are heterogeneous across individuals. In addition, identifying neurobiological underpinnings of behavioral and cognitive abnormalities in ASD, which are needed for rational drug design, has been challenging. Mounting evidence indicates that sensory processing impairments are a key feature of ASD, and aberrant sensory reactivity is now recognized as a core diagnostic symptom (DSM-V, 2013). Furthermore, regression modeling indicates that sensory over-responsivity is strongly correlated with anxiety and gastrointestinal dysfunction, which are frequently co-morbid in people with ASD (Mazurek et al., 2013). Of particular interest is an emerging body of literature indicating that abnormal responses to touch are highly correlated with—and predictive of—ASD severity. Children with ASD often exhibit greater sensitivity to light touch than children with other developmental disorders (Wiggins et al., 2009), and touch avoidance during infancy



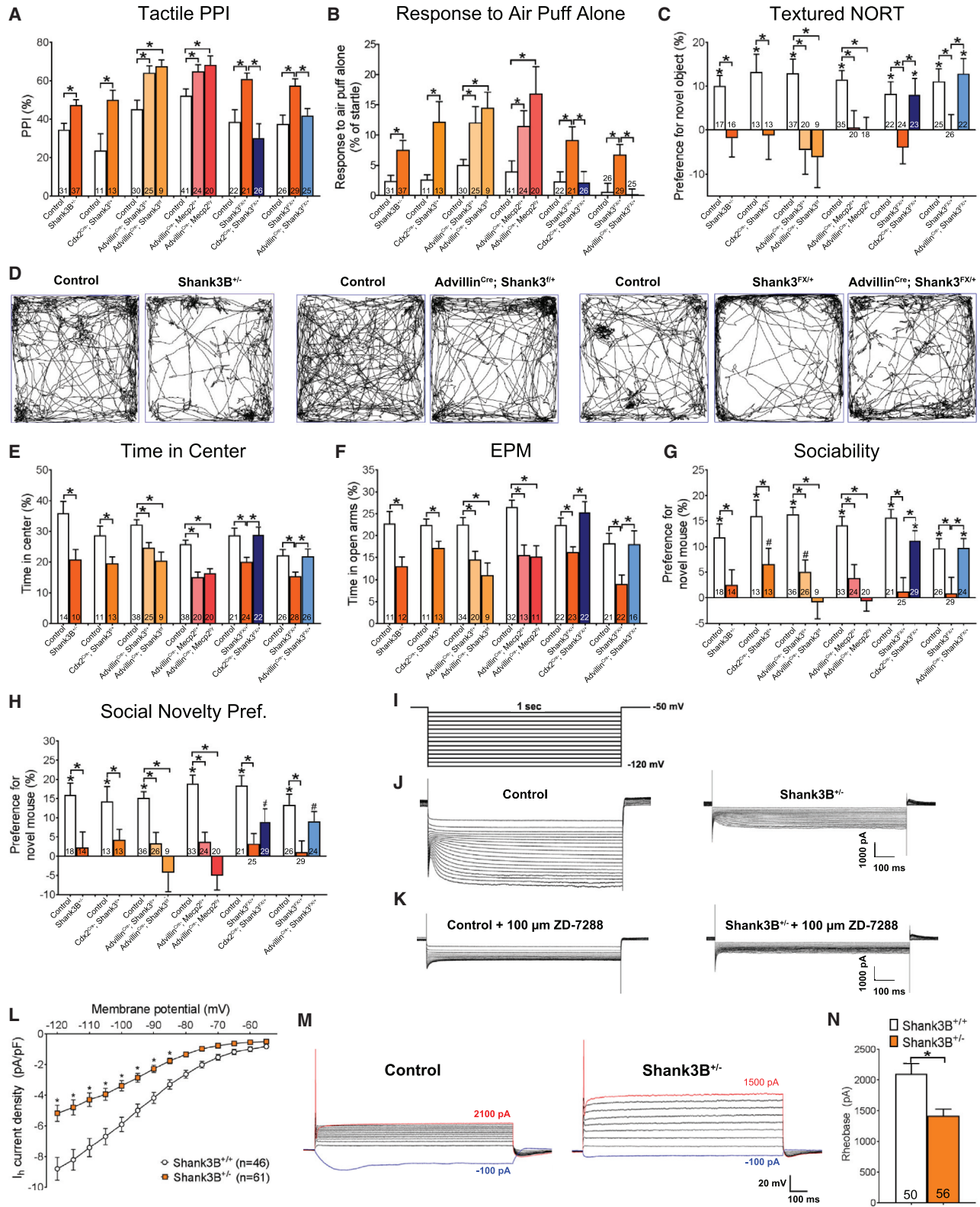


Figure 1. Shank3 Functions Cell-Autonomously in Peripheral Somatosensory Neurons for Normal Innocuous Touch Behaviors

(A) Hairy skin sensitivity was measured using tactile PPI. Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff (250 ms ISI). Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.

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predicts deficits in social development and ASD diagnosis in young children (Mammen et al., 2015).

Recent work aimed at identifying the neurobiological basis of abnormal tactile sensitivity in ASD revealed that peripheral mechanosensory neurons, called low-threshold mechanoreceptor neurons (LTMRs), and their connections within the spinal cord (SC), are dysfunctional in *Mecp2* and *Gabbr3* ASD mouse models due to a loss of GABA_A receptor (GABA_AR)-dependent presynaptic inhibition (PSI) (Orefice et al., 2016). Moreover, aberrant tactile reactivity observed in both *Mecp2* and *Gabbr3* mutant mice contributes to a subset of ASD-associated behavioral phenotypes, including social impairments and anxiety-like behaviors (Orefice et al., 2016). It remains unclear, however, whether peripheral mechanosensory neuron abnormalities are common across additional pervasive developmental disorders associated with ASD, as well as idiopathic ASD. Furthermore, it is not known whether pharmacological strategies targeting peripheral mechanosensory neurons can be used to treat tactile over-reactivity and other ASD-related behaviors. Here, we report that tactile over-reactivity in different ASD models can result from disparate molecular and pathophysiological mechanisms, aberrant tactile reactivity leads to region-specific abnormal brain development, and a peripherally restricted pharmacological approach to suppress tactile over-reactivity during early postnatal development has the potential to improve some behavioral abnormalities associated with ASD.

RESULTS

Shank3 Functions in Peripheral Sensory Neurons for Normal Touch Behaviors and Some ASD-Related Behaviors

We previously found that *Mecp2* and *Gabbr3* function in peripheral somatosensory neurons for normal tactile behaviors (Orefice et al., 2016), however, the extent to which dysfunction of the peripheral nervous system (PNS) neurons contributes to altered somatosensation in other genetic models for ASD is not known. Haploinsufficiency of the gene *Shank3* causes

Phelan-McDermid syndrome, a neurodevelopmental disorder very often presenting with ASD symptoms (Phelan and McDermid, 2012). Patients with Phelan-McDermid syndrome often exhibit enhanced sensitivity to light touch stimuli and tactile defensiveness and, paradoxically, reduced responsivity to certain painful stimuli (Phelan and McDermid, 2012; Philippe et al., 2008). Mice with a germline loss-of-function mutation in *Shank3* (*Shank3B^{+/-}*) exhibit ASD-like traits (Jaramillo et al., 2017; Peça et al., 2011), including tactile over-reactivity (Orefice et al., 2016). SHANK3 is a synaptic protein that is expressed in both mouse and human dorsal root ganglion (DRG) neurons (Ray et al., 2018; Usoskin et al., 2015), including at the presynaptic terminals of LTMRs that are responsible for transmitting light touch information to the SC dorsal horn (Figure S1A). We therefore asked whether peripheral mechanosensory neuron dysfunction in *Shank3* mutant mice underlies abnormal innocuous touch behaviors. To address this, we generated a mouse line harboring a *Shank3* floxed (*Shank3^f*) allele (Figures S1B–S1D) to enable selective ablation of *Shank3* in cells expressing Cre recombinase. *Shank3^f* mice were crossed with mice expressing Cre recombinase below cervical level 2 (*Cdx2^{Cre}*; *Shank3^{f/+}*) (Akyol et al., 2008) or all DRG, trigeminal, and sympathetic ganglia neurons (*Advillin^{Cre}*; *Shank3^{f/+}* or *f/f*) (Hasegawa et al., 2007) (Figure S1A). Of note, *Shank3* is expressed at almost undetectable levels in sympathetic neurons (Furlan et al., 2016). These conditional mutants as well as mice with heterozygous germline *Shank3* deletion (*Shank3B^{+/-}*) and, for comparison, mice with sensory-neuron-specific deletion of *Mecp2* (*Advillin^{Cre}*; *Mecp2^{f/+}* or *f/y*), were subjected to behavioral assays to assess tactile sensitivity and ASD-related behaviors. Consistent with previous findings, *Shank3B^{+/-}* and *Advillin^{Cre}*; *Mecp2^{f/+}* or *f/y* mutant mice exhibited hairy skin hypersensitivity, measured by a tactile prepulse inhibition of an acoustic startle response assay (tactile PPI) and responsivity to an air puff stimulus alone delivered to back hairy skin (Figures 1A and 1B; Table S1) (Orefice et al., 2016). *Shank3B^{+/-}* and *Advillin^{Cre}*; *Mecp2^{f/+}* or *f/y* mutant mice also displayed texture discrimination deficits, assessed using a textured novel object recognition test

(B) Response to a light air puff stimulus alone directed to the back hairy skin. Responses are expressed as percent of startle response to a 125 dB noise. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05.

(C) Texture discrimination was measured using the textured NORT behavioral assay. A positive value indicates preference for the novel object, compared to the familiar object. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05.

(D) Open field (OF) test was used as a general measure of exploration and anxiety-like behavior. Shown are representative activity traces in the OF test for mutant mice and control littermates. OF chamber is 40 cm by 40 cm.

(E) Percent time spent in the center of the OF chamber. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05.

(F) Percent time spent in the open arms of the EPM. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05.

(G) Preference index for the percentage of time spent investigating a novel mouse, compared to a novel object, in the "Sociability" portion of the 3-chamber social interaction test. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05; #*p* < 0.10.

(H) Preference index for the percentage of time spent investigating a novel mouse, compared to a familiar mouse, in the "Social Novelty Preference" portion of the 3-chamber social interaction test. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05; #*p* < 0.10.

(I) Voltage step protocol used to activate HCN channels and elicit *I_h* during whole-cell voltage clamp recordings.

(J and K) Representative electrophysiological traces showing *I_h* during a hyperpolarizing voltage step protocol in large diameter DRG neurons cultured from control and *Shank3B^{+/-}* mutant mice, at baseline (J) and with a selective HCN-channel blocker, ZD-7288 (K).

(L) Quantification of *I_h* density at each voltage step for large diameter neurons cultured from DRGs of control and mutant mice. Two-way ANOVA with post hoc Sidak's test, (F[1,1,470] = 187.7; *p* < 0.0001) **p* < 0.05.

(M) Representative traces from large diameter DRG neurons cultured from control and *Shank3B^{+/-}* mutant mice during whole cell current clamp recordings, in which the minimal amount of current required to elicit an action potential in each neuron (rheobase [*R_h*]), was determined.

(N) Quantification of average *R_h* in large diameter DRG neurons cultured from control and *Shank3B^{+/-}* mutant mice. Student's unpaired t test, **p* < 0.005.

Error bars indicate standard error of the mean. See also Figures S1 and S2 and Tables S1, S2, and S7.

("textured NORT") (Figure 1C) (Orefice et al., 2016). Loss of *Shank3* in cells below the neck (*Cdx2*^{Cre}; *Shank3*^{f/+}) or in somatosensory neurons (*Advillin*^{Cre}; *Shank3*^{f/+} or *f/f*) also led to an increase in tactile PPI, increased responsivity to an air puff stimulus, and deficits in texture discrimination (Figures 1A–1C). *Cdx2*^{Cre}; *Shank3*^{f/+} and *Advillin*^{Cre}; *Shank3*^{f/+} or *f/f* mutant mice were overtly normal, however, and no differences in gross motor behaviors, acoustic PPI, or memory retention were observed among any of the groups (Figures S1E–S1J).

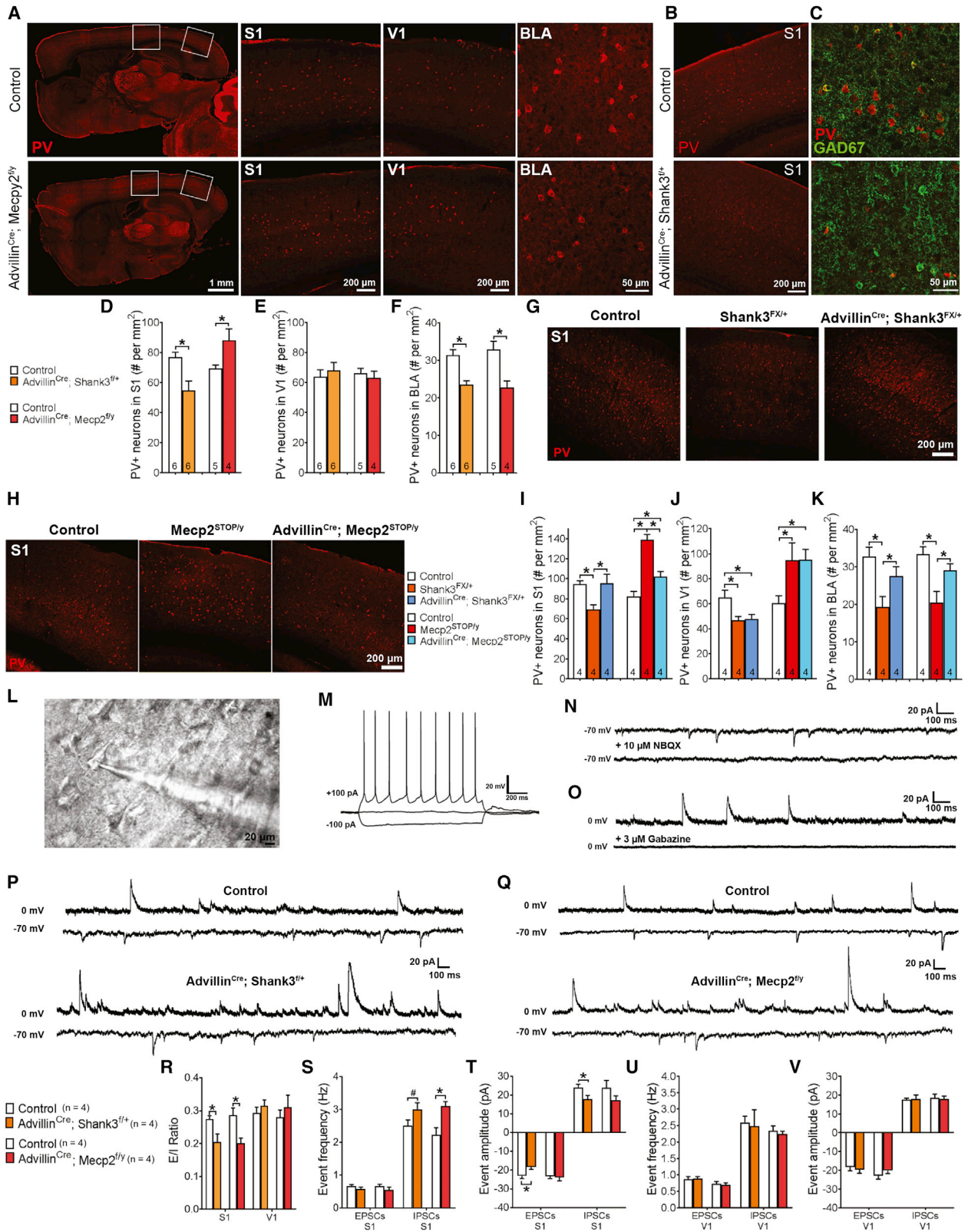
Concomitant with altered reactivity to tactile stimuli, mice in which either *Mecp2* or *Gabrb3* was ablated in peripheral somatosensory neurons during embryonic development exhibited increased anxiety-like behaviors and abnormal social behaviors in adulthood (Figures 1D–1H and S1K–S1M) (Orefice et al., 2016). We therefore hypothesized that altered tactile processing due to developmental loss of *Shank3* in peripheral sensory neurons may also contribute to anxiety-like behaviors and abnormal social behaviors in adult mice. Consistent with prior findings, *Shank3B*^{+/-} mice displayed anxiety-like behaviors in the open field test, elevated plus maze (EPM), and lack of habituation to an acoustic startle noise (Peça et al., 2011) (Figures 1D–1F and S1K–S1M). *Cdx2*^{Cre}; *Shank3*^{f/+}, and *Advillin*^{Cre}; *Shank3*^{f/+} or *f/f* mutant mice also exhibited anxiety-like behaviors (Figures 1D–1F and S1K–S1M). Abnormal social interactions, neophobia, and over-grooming behaviors are additional features of *Shank3B*^{+/-} mutant mice (Jaramillo et al., 2017; Peça et al., 2011). We implemented the three-chamber social interaction test to assess sociability and social novelty recognition preference ("social novelty preference") in the conditional mutants (Silverman et al., 2010). Similar to *Shank3B*^{+/-} mice, the *Cdx2*^{Cre}; *Shank3*^{f/+} and *Advillin*^{Cre}; *Shank3*^{f/+} or *f/f* mice did not show a preference for a novel mouse in either the sociability or social novelty preference portion of the test, although *Cdx2*^{Cre}; *Shank3*^{f/+} and *Advillin*^{Cre}; *Shank3*^{f/+} mutants did exhibit a trend toward preference in the sociability assay (Figures 1G and 1H; Table S2). *Shank3*^{+/-} mice exhibited neophobia during a marble burying task, evidenced by a decreased number of marbles buried compared to controls (Jaramillo et al., 2017) (Figure S1N). A modest decrease in the number of marbles buried was also observed in *Cdx2*^{Cre}; *Shank3*^{f/+} and *Advillin*^{Cre}; *Shank3*^{f/+} or *f/f* mutant mice (Figure S1N). Finally, while *Shank3B*^{+/-} mutants display an overgrooming behavior, this phenotype was not observed in either *Cdx2*^{Cre}; *Shank3*^{f/+} or *Advillin*^{Cre}; *Shank3*^{f/f} mice (Figure S1O). These findings indicate that a subset of behavioral abnormalities observed in *Shank3B*^{+/-} mice are also observed in mice lacking one or both *Shank3* alleles in peripheral somatosensory neurons.

We next asked whether restoration of *Shank3* expression selectively in peripheral somatosensory neurons might improve tactile deficits and other ASD-related phenotypes observed in *Shank3* germline mutant mice. We employed a Cre-dependent genetic switch (FLEX) knockin mouse, which enables conditional expression of *Shank3* from its endogenous genomic locus in the presence of Cre recombinase (Mei et al., 2016) (Figure S1Q). Selective restoration of *Shank3* in cells below the neck (*Cdx2*^{Cre}; *Shank3*^{FX/+}) or in somatosensory neurons (*Advillin*^{Cre}; *Shank3*^{FX/+}) (Figures S1Q and S1R) normalized hairy skin hypersensitivity, textured discrimination deficits and anxiety-like be-

haviors observed in *Shank3*^{FX/+} mice (Figures 1A–1F and S1L). Furthermore, while *Shank3*^{FX/+} mice did not show a preference for a novel mouse in either portion of the 3-chamber social interaction test, both *Cdx2*^{Cre}; *Shank3*^{FX/+} and *Advillin*^{Cre}; *Shank3*^{FX/+} rescue mice exhibited preference for the novel mouse in the sociability assay (Figures 1G and 1H; Table S2). We also observed an intermediate phenotype in the marble burying test: *Cdx2*^{Cre}; *Shank3*^{FX/+} and *Advillin*^{Cre}; *Shank3*^{FX/+} rescue mice buried more marbles than their *Shank3*^{FX/+} littermates, but fewer than control littermates (Figure S1N). Conversely, no improvements in memory deficits or overgrooming behaviors were observed in either *Cdx2*^{Cre}; *Shank3*^{FX/+} or *Advillin*^{Cre}; *Shank3*^{FX/+} rescue mice (Figures S1I, S1O, and S1P). Therefore, as observed for *Mecp2* and *Gabrb3* (Orefice et al., 2016), *Shank3* is required cell-autonomously in peripheral somatosensory neurons for normal tactile sensitivity and texture discrimination as well as a subset of ASD-related behaviors. Thus, dysfunction of mechanosensory neurons is a common feature of multiple ASD mouse models, despite disparate molecular properties and functions of ASD-associated genes.

Distinct Mechanosensory Neuron Physiological Dysfunctions Underlie Altered Tactile Processing across Animal Models for ASD

Prior work indicated that peripheral somatosensory neurons lacking *Mecp2* or *Gabrb3* exhibit decreased expression of GABRB3, the obligatory beta subunit of the GABA_AR in these neurons (Orefice et al., 2016; Zimmerman et al., 2019). This leads to a deficit in GABA_AR-mediated PSI of somatosensory neuron inputs to the SC, thus explaining tactile over-reactivity in *Mecp2* and *Gabrb3* mutants (Orefice et al., 2016). Therefore, we asked whether a similar pathophysiological mechanism could explain tactile over-reactivity observed in *Shank3* mutant mice. However, no alteration in GABRB3 expression in LTMR central terminals of *Shank3* mutant mice was observed (Figures S2E and S2F). Guided by recent observations that *Shank3* ablation in hippocampal neurons causes a reduction of HCN channel expression leading to a loss of the current mediated by these channels, known as *I_h*, as well as increased excitability (Yi et al., 2016), we next asked whether loss of *Shank3* leads to altered expression of HCN channels in sensory neurons. Indeed, fewer HCN1-containing puncta were detected at presynaptic terminals of Aβ- and Aδ-LTMRs in the SC dorsal horn of *Shank3B*^{+/-} and *Advillin*^{Cre}; *Shank3*^{f/+} or *f/f* mutant mice (Figures S2A and S2B). In addition, HCN1 expression was decreased in large but not small diameter DRG cell bodies (Figures S2C and S2D). Consistent with this, whole-cell patch clamp recordings of large diameter sensory neurons cultured from *Shank3* mutant mice revealed multiple physiological alterations. This included increased input resistance, decreased *I_h* elicited during a hyperpolarizing voltage step protocol, and increased excitability as compared to control littermates (Figures 1I–1N, S2G–S2I, and S2N). No differences in capacitance or cell body size of recorded neurons were observed between groups (Figures S2O and S2P). In contrast to these alterations in large diameter neurons, a majority of which are mechanosensory neurons, no deficits in *I_h* or HCN1 expression were observed in small-diameter DRG neurons from *Shank3* mutant mice, although these neurons



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exhibited a modest reduction in excitability (Figures S2C, S2D, and S2J–S2M). For comparison, similar measurements were made using DRG neurons cultured from *Mecp2* mutant mice. Neither large nor small diameter DRG neurons from *Mecp2* mutant mice exhibited abnormalities in I_h , although there was a trend toward a small increase in excitability of large diameter neurons (Figures S2Q–S2S). Consistent with our findings that large diameter DRG neurons from *Shank3B^{+/-}* mice display decreased I_h and altered excitability, *Hcn1^{-/-}* mice exhibited enhanced hairy skin sensitivity and deficits in texture discrimination (Figures S2T–S2W). Together, these findings indicate that multiple ASD-associated genes function cell-autonomously in peripheral somatosensory neurons for normal tactile sensitivity, and at least two distinct pathophysiological mechanisms underlie tactile over-responsivity in ASD mouse models: loss of GABA_AR-mediated PSI of LTMR inputs to the SC dorsal horn, as observed in *Mecp2* and *Gabrb3* mutants (Orefice et al., 2016), or decreased I_h and hyperexcitability of somatosensory neurons, observed in *Shank3* mutants.

ASD-Related Gene Mutations in Somatosensory Neurons Lead to Region-Specific Alterations in Brain Inhibitory Interneurons and Cortical Microcircuit Properties

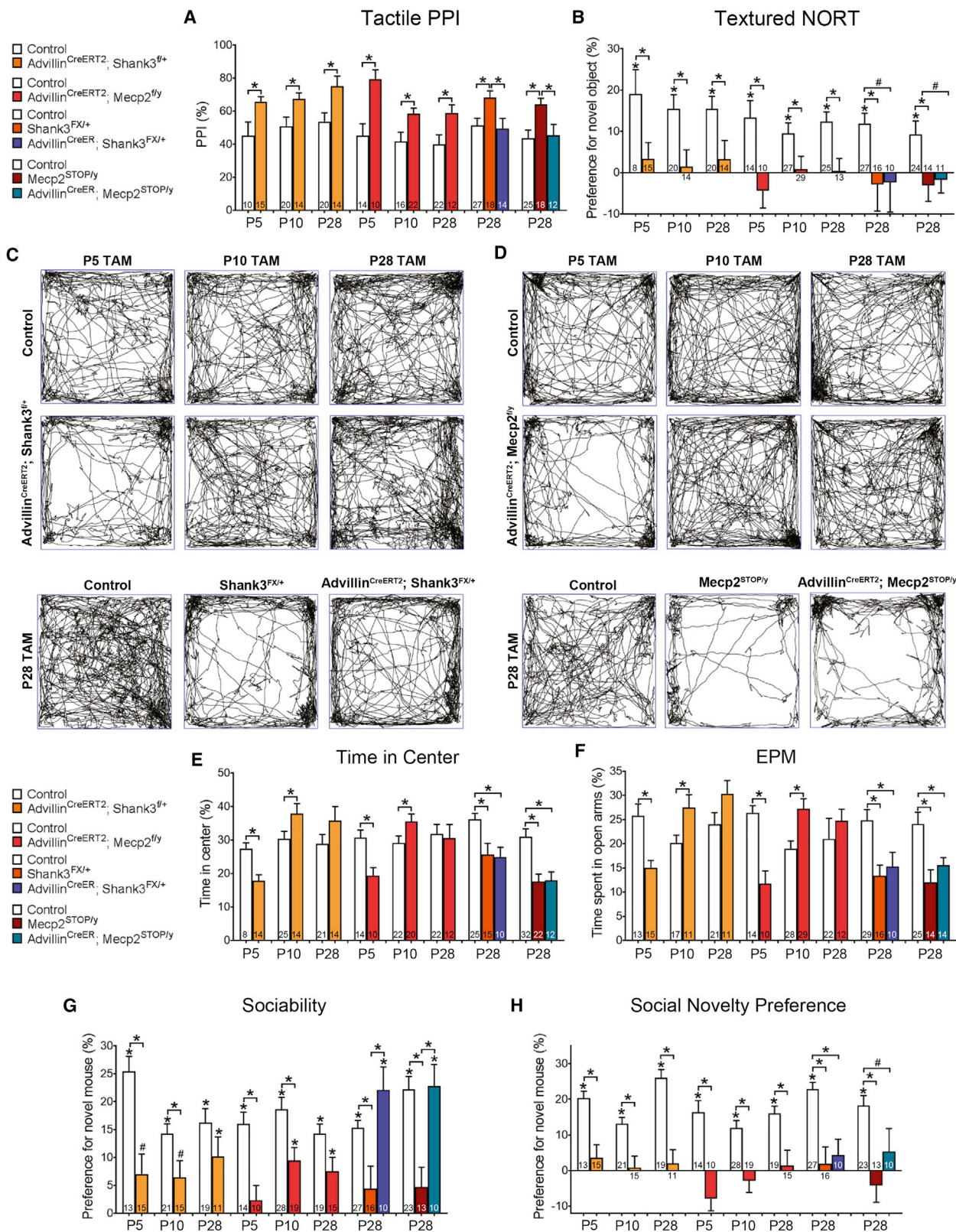
We next explored the relationship between aberrant tactile reactivity, caused by ablation of either *Mecp2* or *Shank3* in sensory neurons, and brain development and cortical microcircuit properties. One clue that may help to explain any potential relationship stems from decades of research indicating that sensory experience guides development of neocortical areas where features of sensory stimuli are represented (Hubel and Wiesel, 1970; Simons and Land, 1987; Wiesel and Hubel, 1965). In line with this, postmortem analysis of brains from ASD patients as well

as animal models for ASD (such as *Mecp2* and *Shank3* mutants) revealed abnormalities in parvalbumin-positive (PV⁺) inhibitory interneurons in multiple brain regions, including primary sensory cortices (Filice et al., 2016; Fukuda et al., 2005; Hashemi et al., 2017; Marín, 2012; Nelson and Valakh, 2015; Tomassy et al., 2014; Zikopoulos and Barbas, 2013). Cortical PV⁺ neurons are fast-spiking interneurons that contribute to feed-forward and feedback inhibition and modulate sensory responsiveness (König et al., 1996; Sohal et al., 2009; Womelsdorf et al., 2014). Therefore, we hypothesized that aberrant sensory inputs caused by peripheral somatosensory neuron dysfunction in ASD models may affect properties of PV⁺ inhibitory interneurons in brain circuits that process these inputs. In agreement with this idea, loss of *Mecp2* in peripheral somatosensory neurons in *Advillin^{Cre}; Mecp2^{f/y}* mice caused an increased number of PV⁺ interneurons in primary somatosensory cortex (S1), but not primary visual cortex (V1), of adult mice compared to controls (Figures 2A, 2D, and 2E; Table S1). However, loss of *Shank3* in sensory neurons, in *Advillin^{Cre}; Shank3^{f/+}* mice, led to a reduction in PV⁺ neurons in S1, but not V1, of adult mice (Figures 2B–2E). These region-specific alterations in PV⁺ interneurons in *Shank3* and *Mecp2* conditional mutants are in contrast to the more widespread alterations observed in the germline mutants: *Mecp2^{STOP/y}* and *Shank3^{FX/+}* germline mutant mice exhibited alterations in PV⁺ neurons in both S1 and V1 (Figures 2G–2J). We also observed a decrease in the number of PV⁺ neurons in the basolateral amygdala (BLA), a region of the brain that plays critical roles in anxiety and social behaviors, in both *Shank3* and *Mecp2* conditional mutants (Figures 2A and 2F). Conversely, developmental restoration of either *Shank3* or *Mecp2* in somatosensory neurons in *Advillin^{Cre}; Shank3^{FX/+}* and *Advillin^{Cre}; Mecp2^{STOP/y}* mice, respectively, improved the PV⁺ neuron abnormalities observed in *Shank3^{FX/+}* and *Mecp2^{STOP/y}* mice in

Figure 2. Loss of Either *Shank3* or *Mecp2* in Peripheral Somatosensory Neurons Leads to Abnormal Brain Interneuron Development and Microcircuit Properties in a Region-Specific Manner

- (A) Representative immunohistochemistry (IHC) images of parvalbumin (PV) immunoreactivity in control or *Advillin^{Cre}; Mecp2^{f/y}* mutant mice, showing full sagittal brain sections and corresponding magnified sections of primary somatosensory cortex (S1) and primary visual cortex (V1), or coronal sections of basolateral amygdala (BLA).
- (B) Representative IHC images of coronal brain sections, showing PV immunoreactivity in control or *Advillin^{Cre}; Shank3^{f/+}* mutant mice.
- (C) Representative IHC images of coronal S1 brain sections, showing PV and GAD67 immunoreactivity in control or *Advillin^{Cre}; Shank3^{f/+}* mutant mice.
- (D–F) Quantification of the number of PV-positive (PV⁺) neurons in S1 (D), V1 (E), and BLA (F) in mutant mice and their control littermates. Student's unpaired t test, *p < 0.05.
- (G) Representative IHC images of coronal S1 brain sections, showing PV immunoreactivity in control, *Shank3^{FX/+}*, or *Advillin^{Cre}; Shank3^{FX/+}* mutant mice.
- (H) Representative IHC images of coronal S1 brain sections, showing PV immunoreactivity in control, *Mecp2^{STOP/y}*, or *Advillin^{Cre}; Mecp2^{STOP/y}* mutant mice.
- (I–K) Quantification of the number of PV⁺ neurons in S1 (I), V1 (J), and BLA (K) in mutant mice and their control littermates. One-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (L) Example image of a coronal S1 slice with a layer 2/3 pyramidal neuron in whole cell patch clamp recording configuration.
- (M) Example firing pattern of a layer 2/3 pyramidal neuron in whole cell patch clamp recording configuration during current injection steps.
- (N) Example traces from a layer 2/3 pyramidal neuron showing spontaneous excitatory postsynaptic currents (sEPSCs) in normal bath solution or following application of NBQX (10 μM) to block AMPA receptors.
- (O) Example traces from a layer 2/3 pyramidal neuron showing spontaneous inhibitory postsynaptic currents (sIPSCs) in normal bath solution or following application of gabazine (3 μM) to block GABA_A receptors.
- (P) Representative traces showing sEPSCs (–70 mV hold) and sIPSCs (0 mV hold) from S1 slices of control or *Advillin^{Cre}; Shank3^{f/+}* mutant mice.
- (Q) Representative traces showing sEPSCs (–70 mV hold) and sIPSCs (0 mV hold) from S1 slices of control or *Advillin^{Cre}; Mecp2^{f/y}* mutant mice.
- (R) Quantification of excitatory/inhibitory (E/I) ratios in S1 or V1 slices from control and mutant mice. Two-way ANOVA with post hoc Sidak's test, *p < 0.05.
- (S and T) Quantification of sEPSC and sIPSC event frequency (S) and event amplitude (T) in S1 slices from control and mutant mice. Two-way ANOVA with post hoc Sidak's test, *p < 0.05; #p < 0.10.
- (U and V) Quantification of sEPSC and sIPSC event frequency (U) and event amplitude (V) in V1 slices from control and mutant mice. Two-way ANOVA with post hoc Sidak's test, *p < 0.05.

Error bars indicate standard error of the mean. See also Tables S1 and S7.



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both S1 and BLA, but not in V1 (Figures 2G–2K). Related to this, we also found that loss of either *Mecp2* or *Shank3* in somatosensory neurons led to decreased excitatory/inhibitory (E/I) ratios measured electrophysiologically in layer 2/3 pyramidal neurons using acute S1 slices, compared to control littermates (Figure 2R). Layer 2/3 pyramidal neurons from S1 slices of *Advillin^{Cre2}*; *Mecp2^{f/y}* mice exhibited an increase in spontaneous inhibitory postsynaptic current (sIPSC) frequency, with no differences observed in spontaneous excitatory postsynaptic current (sEPSC) or sIPSC amplitudes (Figures 2Q, 2S, and 2T). On the other hand, while layer 2/3 pyramidal neurons from S1 slices of *Advillin^{Cre}*; *Shank3^{f/+}* mice showed no difference in the frequency of sIPSCs, decreased sEPSC and sIPSC amplitudes were observed in these mutants (Figures 2P, 2S, and 2T). No differences in either the amplitude or frequency of events were observed in V1 in any of the conditional mutants analyzed (Figures 2U and 2V). These analyses indicate that physiological dysfunction of peripheral somatosensory neurons caused by distinct ASD-associated gene mutations leads to functional changes in cortical microcircuit properties in a region-specific manner.

A Critical Window during which Somatosensory Neuron Dysfunction Influences Cognitive and Social Behaviors

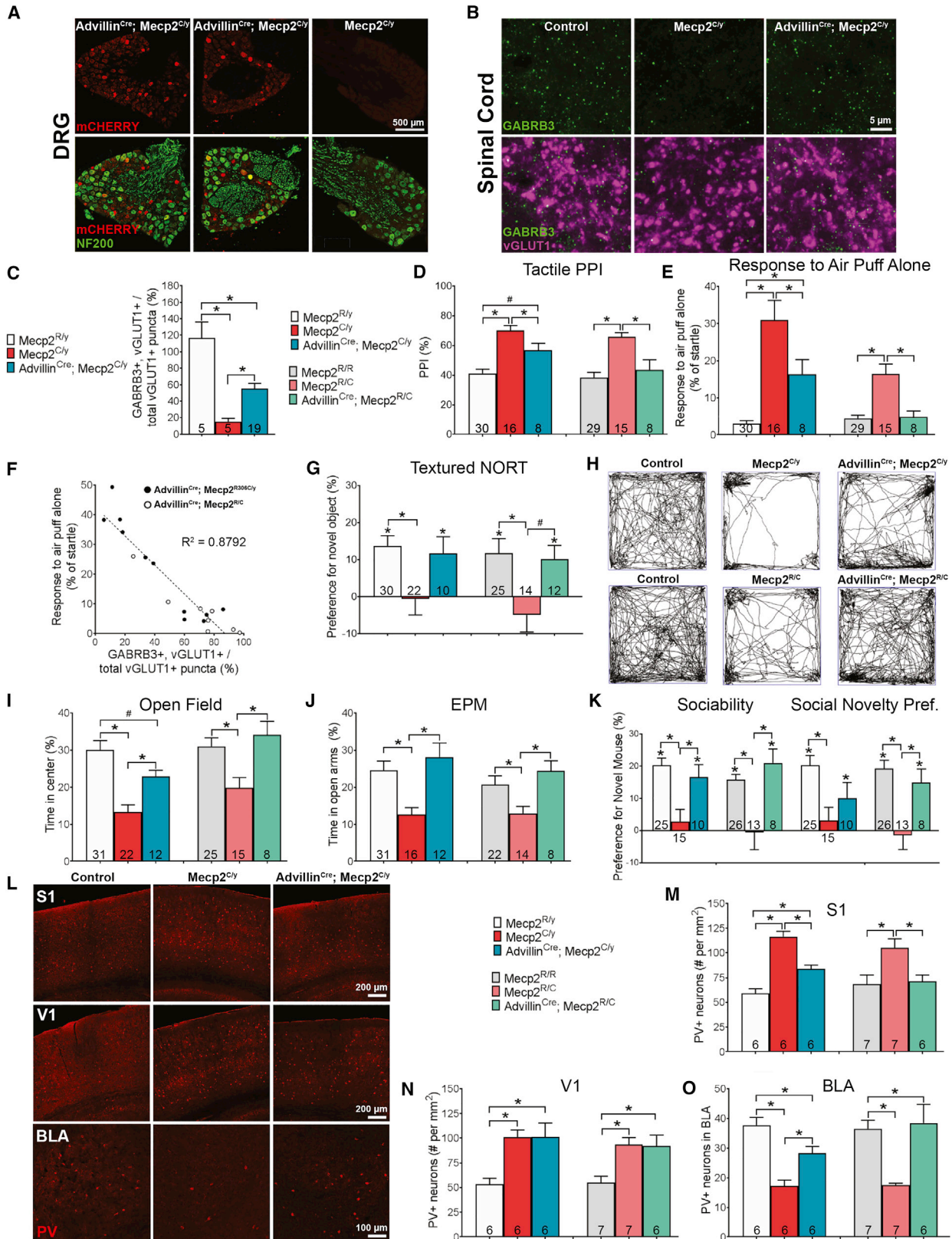
We next determined when, during development, ASD-associated genes function in peripheral somatosensory neurons to govern tactile reactivity, brain microcircuit development, and behavior. To address this, we used the tamoxifen-sensitive *Advillin^{CreERT2}* mouse line (Lau et al., 2011) to delete either *Shank3* or *Mecp2* during different postnatal developmental stages (Figures S4A–S4D) and then assessed tactile reactivity, texture discrimination, PV⁺ interneurons in S1, V1 and BLA, and cognitive and social behaviors in young adult mice beginning at 6 weeks of age. Early postnatal deletion of either *Shank3* or *Mecp2*, beginning at P5 (*Advillin^{CreERT2}*; *Shank3^{f/+}* or *Advillin^{CreERT2}*; *Mecp2^{f/y}*), recapitulated the behavioral phenotypes observed in *Advillin^{Cre}*; *Shank3^{f/+}* and *Advillin^{Cre}*; *Mecp2^{f/y}* mice (Figures 3, S3A, and S3H–S3K; Tables S3 and S4). Moreover, similar to *Advillin^{Cre}*; *Mecp2^{f/y}* mice, *Advillin^{CreERT2}*; *Mecp2^{f/y}* mutant mice receiving P5 tamoxifen treatment exhibited more PV⁺ neurons in S1 (Figure S4G). Likewise, *Advillin^{CreERT2}*; *Shank3^{f/+}* mutant mice displayed fewer PV⁺ neurons in S1, which was similar to that observed in *Advillin^{Cre}*; *Shank3^{f/+}* mice (Figure S4G). Decreased density of PV⁺ neurons in BLA was also observed in both *Advillin^{CreERT2}*; *Shank3^{f/+}* and *Advillin^{CreERT2}*; *Mecp2^{f/y}* mutant mice receiving

the P5 tamoxifen treatment (Figure S4I). In contrast, while deletion of either *Mecp2* or *Shank3* in somatosensory neurons (*Advillin^{CreERT2}*; *Mecp2^{f/y}* or *Advillin^{CreERT2}*; *Shank3^{f/+}*) at P28 led to abnormalities in tactile behaviors and PV⁺ neuron density in S1, these mice did not exhibit anxiety-like behaviors or neophobia, and their social impairments were considerably less severe than mice with embryonic or P5 deletion of either gene in peripheral sensory neurons (Figures 3, S3A, S3H–S3K, S4F, and S4G; Table S3). Mice with P28 deletion also displayed no abnormalities in BLA PV⁺ neuron density (Figure S4I). Interestingly, when either *Mecp2* or *Shank3* was ablated in somatosensory neurons beginning at an intermediate time point, P10, mice exhibited a distinct set of behavioral alterations. In P10 deletions, tactile and social behavior abnormalities were observed, similar to embryonic deletions, but this was accompanied by hyperactivity and reduced anxiety-like behaviors (Figures 3, S3A, and S3H–S3M; Table S3). Moreover, P10 ablations of *Shank3* and *Mecp2* in somatosensory neurons both lead to a reduction in PV⁺ neurons in S1 but increased density of PV⁺ neurons in the BLA (Figures S4G–S4I).

In related experiments, we asked whether postnatal restoration of *Mecp2* or *Shank3* function in somatosensory neurons might improve ASD-related phenotypes in mice. For this, *Advillin^{CreERT2}* mice crossed to either *Shank3^{FX/+}* or *Mecp2^{STOP/+}* mice were used to restore *Shank3* or *Mecp2* expression, respectively, in somatosensory neurons following 5 days of tamoxifen administration beginning at P28 (Figure S4J). Here, restoration of either *Shank3* or *Mecp2* in somatosensory neurons, beginning at P28, was sufficient to normalize hairy skin hypersensitivity observed in *Shank3^{FX/+}* or *Mecp2^{STOP/y}* littermates (Figures 3A and S3A). However, memory deficits were not improved in either *Advillin^{CreERT2}*; *Shank3^{FX/+}* or *Advillin^{CreERT2}*; *Mecp2^{STOP/y}* P28 rescue mice (Figure S3F), and motor impairments typically observed in *Mecp2* germline mutant mice were also not improved in *Advillin^{CreERT2}*; *Mecp2^{STOP/y}* mice (Figure S3B). Furthermore, *Advillin^{CreERT2}*; *Shank3^{FX/+}* and *Advillin^{CreERT2}*; *Mecp2^{STOP/y}* mice treated with tamoxifen beginning at P28 did not show improvements in texture discrimination deficits, anxiety-like behaviors or neophobia (Figures 3B–3F and S3B–S3I). *Advillin^{CreERT2}*; *Shank3^{FX/+}* and *Advillin^{CreERT2}*; *Mecp2^{STOP/y}* rescue mice showed modest improvements in social behaviors, with a significant preference for a novel mouse in the sociability assay, but not in the social novelty preference test (Figures 3G and 3H; Table S3). These findings indicate that normal tactile reactivity is necessary during postnatal development for the acquisition of normal brain microcircuit properties and cognitive behaviors.

Figure 3. Normal Tactile Sensitivity Is Necessary during Early Postnatal Periods for Normal Brain Development and Behavior

(A) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in mutant mice and control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05. For experiments shown in this figure, tamoxifen was administered to all littermates over a 5-day period beginning on the day noted for each condition. For all behavioral experiments in this figure, animals began testing at P42. (B) Discrimination index for textured NORT. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05. (C) Representative activity traces in the OF test for *Shank3* mutant mice and control littermates. Tamoxifen (TAM) was administered over a 5-day period, beginning on the day noted for each condition. OF chamber is 40 cm by 40 cm. (D) Representative activity traces in the OF test for *Mecp2* mutant mice and control littermates. (E) Percent time spent in the center of the OF chamber. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05. (F) Percent time spent in the open arms of the EPM. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05. (G and H) Preference index for the percentage of time spent investigating the novel mouse in the "Sociability" (G) and "Social Novelty Preference" (H) portions of the 3-chamber social interaction test. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, **p* < 0.05; #*p* < 0.10. Error bars indicate standard error of the mean. See also Figures S3 and S4 and Tables S3, S4, and S7.



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Postnatal Viral Restoration of GABRB3 Improves Behavioral Deficits in an *Mecp2* Mouse Model of RTT/ASD

Our findings suggest that targeting the PNS may provide an opportunity for improving tactile reactivity, anxiety-like behaviors, and some social behaviors in adult mice harboring ASD-associated gene mutations, with optimal intervention occurring early during postnatal life. Therefore, we considered whether enhancing GABA_AR signaling in peripheral sensory neurons may provide a means of attenuating tactile over-reactivity in ASD models and, in doing so, improve anxiety-like behaviors and potentially other ASD-associated behaviors. The rationale for this idea stems from the finding that *Mecp2* mutant mice exhibit decreased expression of the GABA_AR obligatory subunit GABRB3 at presynaptic terminals of LTMRs in the SC, leading to functional deficits in PSI of mechanosensory neuron input to the SC and altered tactile processing (Orefice et al., 2016). Using conditional mouse genetics and a FLEEx AAV delivery strategy, we were able to selectively restore GABRB3 expression in somatosensory neurons of *Mecp2* mutant mice in a temporally precise manner. Mice harboring an arginine-to-cysteine missense mutation in *Mecp2* (*Mecp2*^{R306C}), which is a common allele found in patients with Rett syndrome (RTT) (Lyst et al., 2013), were used in these studies. We used *Advillin*^{Cre} mice and intraperitoneal (i.p.) injection of an AAV9.FLEEx.GABRB3.mCHERRY virus at P5, to selectively express GABRB3 in peripheral sensory neurons. Thus, DRG neurons in mice harboring the *Advillin*^{Cre} allele expressed functional GABRB3 following i.p. injection of P5 pups with AAV9.FLEEx.GABRB3.mCHERRY (Figures 4A–4C, S5A, and S5B). Increased expression of GABRB3 in somatosensory neurons improved tactile behavior abnormalities in a dose-dependent manner: *Advillin*^{Cre}; *Mecp2*^{C/y} mice displayed significant improvements in hairy skin hypersensitivity and texture

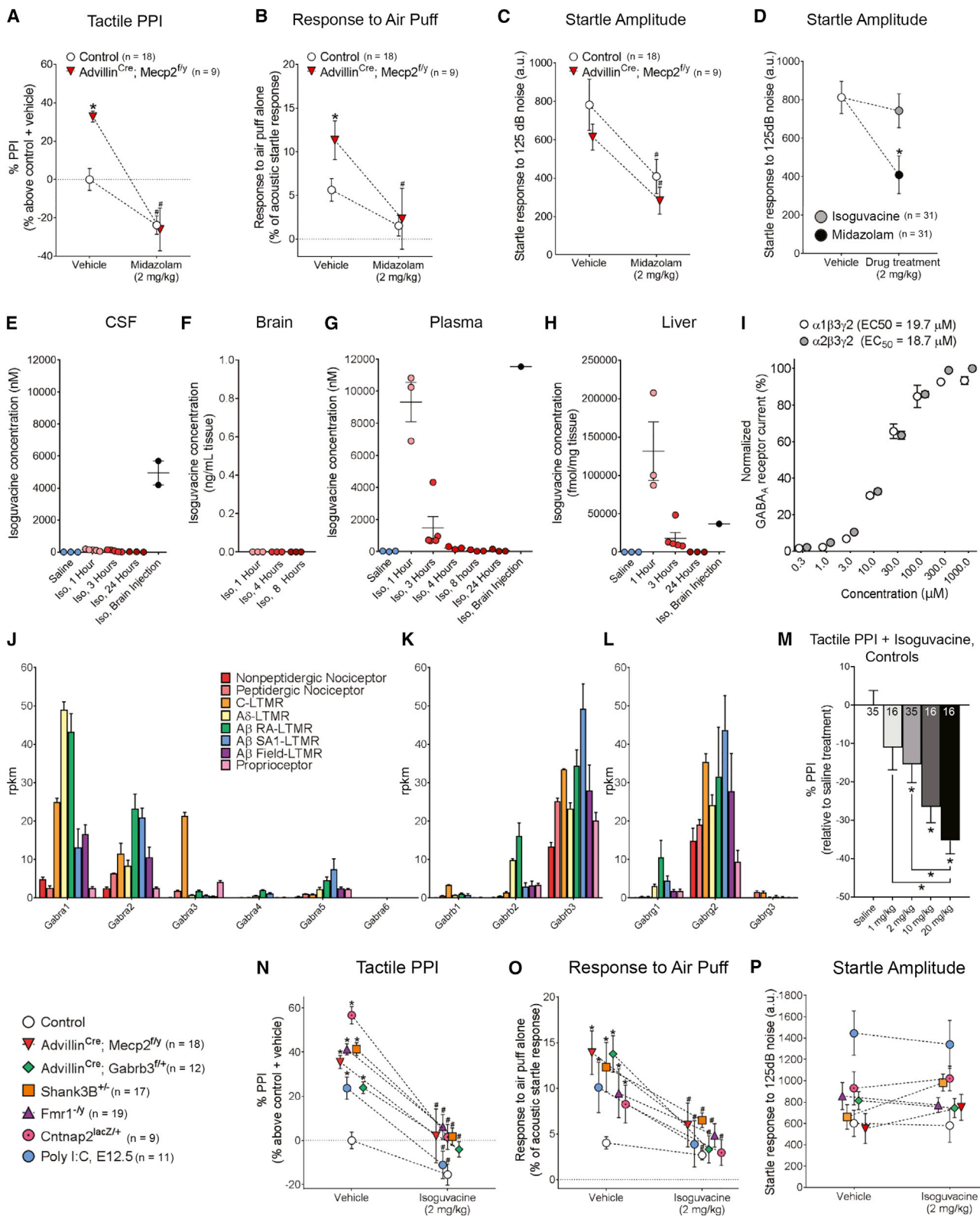
discrimination, compared to *Mecp2*^{C/y} mice (Figures 4D–4G; Table S5). Female “GABRB3 rescue” mice, which are heterozygous for the *Mecp2* mutation (*Advillin*^{Cre}; *Mecp2*^{R/C}), showed a normalization of somatosensory phenotypes (Figures 4D–4G). Similarly, we found a significant correlation between both the number of DRG neurons transduced and the number of GABRB3 puncta in the dorsal horn and reduced responsivity to an air puff stimulus in *Mecp2* mutants (Figures 4F and S5H). Restoration of GABRB3 expression in somatosensory neurons also improved anxiety-like behaviors, social impairments, and PV⁺ neuron abnormalities in S1 and BLA (but not V1), of both male and female rescue mice, with female *Advillin*^{Cre}; *Mecp2*^{R/C} mice exhibiting values similar to those of control littermates (Figures 4H–4O and S5L–S5P). Augmented GABRB3 expression, however, did not improve memory deficits, motor impairments or heightened acoustic PPI performance in mutant animals (Figures S5C–S5E, S5I, and S5J). This proof-of-concept experiment suggests that increasing GABA_AR function in somatosensory neurons during early postnatal development ameliorates tactile over-reactivity, altered S1 and BLA microcircuits, and at least some behavioral deficits observed in germline *Mecp2* mutants.

Acute Treatment with a Peripherally Restricted GABA_AR Agonist Improves Hairy Skin Over-Reactivity in Six Distinct, Genetic, and Environmental Models for ASD

Results of the GABA_AR restoration experiments raised the possibility that pharmacological treatment with a GABA_AR agonist may improve tactile reactivity in *Mecp2* mutant mice. We therefore tested whether benzodiazepines, which are positive allosteric modulators (PAMs) of GABA_ARs, would attenuate hairy skin over-reactivity in *Mecp2* mutant mice. As expected, *Advillin*^{Cre}; *Mecp2*^{f/y} mice receiving vehicle treatment showed tactile hypersensitivity compared to control littermates (Figures 5A and

Figure 4. Viral Expression of GABRB3 in Peripheral Sensory Neurons, Beginning at P5, Improves Some Tactile, Brain, and Behavioral Deficits Observed in *Mecp2*^{R306C} Mutant Mice

- (A) Intraperitoneal (i.p.) injection of AAV.FLEEx.GABRB3.mCHERRY into *Advillin*^{Cre}; *Mecp2*^{C/y} hemizygous null mutant mice at P5 transduces peripheral sensory neurons, as evidenced by immunoreactivity for mCHERRY in two example IHC images of transverse DRG sections from *Advillin*^{Cre}; *Mecp2*^{C/y} mice. Transduced large diameter neurons are immunoreactive for both mCHERRY and NF200. Note lack of mCHERRY immunoreactivity in example image of a transverse DRG section from a *Mecp2*^{C/y} mouse. All littermates received i.p. injection of AAV.FLEEx.GABRB3.mCHERRY at P5.
- (B) IHC images of spinal cord (SC) dorsal horn lamina III/IV from male control, *Mecp2*^{C/y}, or *Advillin*^{Cre}; *Mecp2*^{C/y} mice, showing GABRB3 puncta at vGLUT1⁺ presynaptic terminals for A β and A δ LTMRs.
- (C) Quantification of vGLUT1⁺ puncta co-labeled with GABRB3, relative to the total number of vGLUT1⁺ puncta visualized per image of SC dorsal horn. One-way ANOVA with post hoc Tukey's test, **p* < 0.01.
- (D) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in male control, *Mecp2*^{C/y}, *Advillin*^{Cre}; *Mecp2*^{C/y} or female control, *Mecp2*^{R/C}, and *Advillin*^{Cre}; *Mecp2*^{R/C} mice. One-way ANOVA with post hoc Tukey's test, **p* < 0.05; #*p* < 0.10.
- (E) Response to a light air puff stimulus alone. Responses are expressed as percent of startle response to a 125 dB noise. One-way ANOVA with post hoc Tukey's test, **p* < 0.05.
- (F) The percentage of vGLUT1⁺ puncta co-labeled with GABRB3, relative to the total number of vGLUT1⁺ puncta is negatively correlated with hairy skin sensitivity. Comparison of individual animals' expression levels of GABRB3 at vGLUT1⁺ terminals to their responses to a light air puff stimulus (50 ms, 0.9 PSI). Linear regression analysis [F(1,17) = 123.8, *P* < 0.0001], *R*² = 0.8792.
- (G) Discrimination index for textured NORT. One-way ANOVA with post hoc Tukey's test, **p* < 0.05; #*p* < 0.10.
- (H) Representative activity traces in the OF test. OF chamber is 40 cm by 40 cm.
- (I) Percent time spent in the center of the OF chamber. One-way ANOVA with post hoc Tukey's test, **p* < 0.05; #*p* < 0.10.
- (J) Percent time spent in the open arms of the EPM. One-way ANOVA with post hoc Tukey's test, **p* < 0.05.
- (K) Preference index for the percentage of time spent investigating the novel mouse in the “Sociability” or “Social Novelty Preference” portion of the 3-chamber social interaction test. One-way ANOVA with post hoc Tukey's test, **p* < 0.05.
- (L) Representative IHC images of coronal S1, V1 and BLA brain sections, showing PV immunoreactivity in control, *Mecp2*^{C/y}, or *Advillin*^{Cre}; *Mecp2*^{C/y} mice.
- (M–O) Quantification of the number of PV⁺ neurons in S1 (M), V1 (N), and BLA (O) in mutants, mutant rescues, and their control littermates. One-way ANOVA with post hoc Tukey's test, **p* < 0.05.
- Error bars indicate standard error of the mean. See also Figure S5 and Tables S5 and S7.



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5B; Table S5). Midazolam treatment attenuated tactile sensitivity in both *Mecp2* mutants and controls (Figures 5A and 5B). However, this treatment also caused significant sedation in all mice tested (Figure 5C), complicating interpretations and limiting the utility of this approach. In line with this, traditional blood-brain barrier (BBB) penetrating GABA_AR PAMs are useful for treating some ASD-related symptoms in patients, including acute anxiety, but adverse effects such as sedation and addictive potential are problematic (Gudex, 1991). Furthermore, long-term benzodiazepine use is linked to cognitive impairment across multiple domains, including short-term verbal memory and attention (Golombok et al., 1988; Tata et al., 1994). These considerations prompted us to ask whether peripherally restricted GABA_AR agonists administered beginning at a young age could ameliorate tactile hypersensitivity in ASD models with fewer adverse effects than brain-penetrating benzodiazepines, including sedation and cognitive impairment, which are attributed to a CNS site of drug action. Isoguvacine is a potent and selective agonist for the GABA_AR (Bowery et al., 1983; Hill and Bowery, 1981; Krogsgaard-Larsen and Johnston, 1978; Krogsgaard-Larsen et al., 1977). Due to its zwitterionic structure, isoguvacine is predicted to not cross the BBB (Krogsgaard-Larsen et al., 1981). In agreement with this, we found that isoguvacine administration (2 mg/kg, i.p.) did not cause sedation in mice, which contrasts with BBB-penetrating benzodiazepine midazolam treatment (Figure 5D). Moreover, bio-distribution analyses indicated that following i.p. administration of isoguvacine (20 mg/kg) the drug was detected in blood and liver, but was undetectable in the

brain and cerebrospinal fluid, confirming that isoguvacine does not cross the BBB (Figures 5E–5H). RNA deep sequence analysis of genetically labeled DRG neuron subtypes (Zheng et al., 2019) revealed that $\alpha 1\beta 3\gamma 2$ - and $\alpha 2\beta 3\gamma 2$ -containing GABA_ARs are the major holo-pentameric GABA_ARs expressed in LTMR subtypes, with lower levels of expression in nociceptors and proprioceptors (Figures 5J–5L and S6H). Using a GABA_AR chloride flux assay, isoguvacine was observed to potently activate both $\alpha 1\beta 3\gamma 2$ - and $\alpha 2\beta 3\gamma 2$ -containing GABA_ARs, with EC₅₀ values of 19.7 μ M and 18.7 μ M, respectively (Figure 5I).

We found that acute administration of isoguvacine (2 mg/kg, i.p.) reduced tactile reactivity in control, *Advillin*^{Cre}, *Mecp2*^{f/y} and *Advillin*^{Cre}; *Gabrb3*^{f/+} mutant mice (Figures 5M–5O). These findings prompted us to ask whether this peripherally restricted GABA_AR agonist may attenuate tactile over-reactivity in other ASD mouse models regardless of the pathophysiological mechanism of mechanosensory neuron dysfunction. Indeed, acute administration of isoguvacine also attenuated hairy skin hypersensitivity in *Shank3*, *Fmr1*, and *Cntnap2* mutant mice (Figures 5N and 5O) as well as mice exposed to polyinosinic:polycytidylic acid (Poly I:C) at E12.5 (Figures 5N and 5O), which is a model of maternal immune activation-induced ASD in rodents (Choi et al., 2016). Of note, hairy skin hyposensitivity was observed in 16p11.2 deletion mutant mice (*16p11.2*^{+/-}), and no alterations in tactile behaviors were observed in mice with a deletion spanning exons 4–9 in *Shank3* (*Shank3*^{ex4-9}) (Figures S6I–S6N). Isoguvacine administration led to a further reduction in tactile PPI in *16p11.2*^{+/-} mice (Figure S6I). Consistent with a peripheral

Figure 5. Acute Administration of the Peripherally Restricted GABA_A Receptor Agonist Isoguvacine Improves Tactile Hypersensitivity in Five Genetic and One Environmental Model for ASD

(A) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in mice following i.p. administration of either saline or 2 mg/kg midazolam treatment. Repeated-measures, two-way ANOVA with post hoc Sidak's test, **p* < 0.05 for comparisons between saline-treated mutants and saline-treated control littermates; #*p* < 0.05, for comparisons between saline and isoguvacine treatments within the same genotype.

(B) Response to a light air puff stimulus alone in mice following i.p. administration of either saline or 2 mg/kg midazolam treatment. Responses are expressed as percent of startle response to a 125 dB noise. Repeated-measures, two-way ANOVA with post hoc Sidak's test, **p* < 0.05 for comparisons between saline-treated mutants and saline-treated control littermates; #*p* < 0.05, for comparisons between saline and isoguvacine treatments within the same genotype.

(C) Magnitude of startle response to a 125 dB noise in mice following i.p. administration of either saline or 2 mg/kg midazolam treatment. Repeated-measures, two-way ANOVA with post hoc Sidak's test, **p* < 0.05 for comparisons between saline-treated mutants and saline-treated control littermates; #*p* < 0.05, for comparisons between saline and isoguvacine treatments within the same genotype.

(D) Magnitude of startle response to a 125 dB noise in control mice following i.p. administration of saline, midazolam (2 mg/kg) or a peripherally restricted GABA_A receptor agonist, isoguvacine (2 mg/kg). Repeated-measures, two-way ANOVA with post hoc Sidak's test, **p* < 0.05.

(E–H) Liquid chromatography mass spectrometry (LC-MS) quantified isoguvacine concentrations in cerebrospinal fluid (CSF) (E), brain homogenate (F), plasma (G), or liver (H) samples of mice treated with: i.p. saline, i.p. isoguvacine ("Iso," 20 mg/kg, samples collected multiple time points post-injection), or intracerebral injection isoguvacine (20 mg/kg, 1 h post-injection).

(I) Normalized peak chloride flux through $\alpha 1\beta 3\gamma 2$ - and $\alpha 2\beta 3\gamma 2$ -containing GABA_A receptors in response to isoguvacine application. Response data has been normalized to the baseline peak current induced by addition of EC₁₀₀ GABA (30 μ M) for 2 seconds during the assay.

(J–L) GABA_A receptor subunit RNA expression levels across peripheral somatosensory neuron subtypes, for the alpha (J), beta (K), and gamma (L) subunit types.

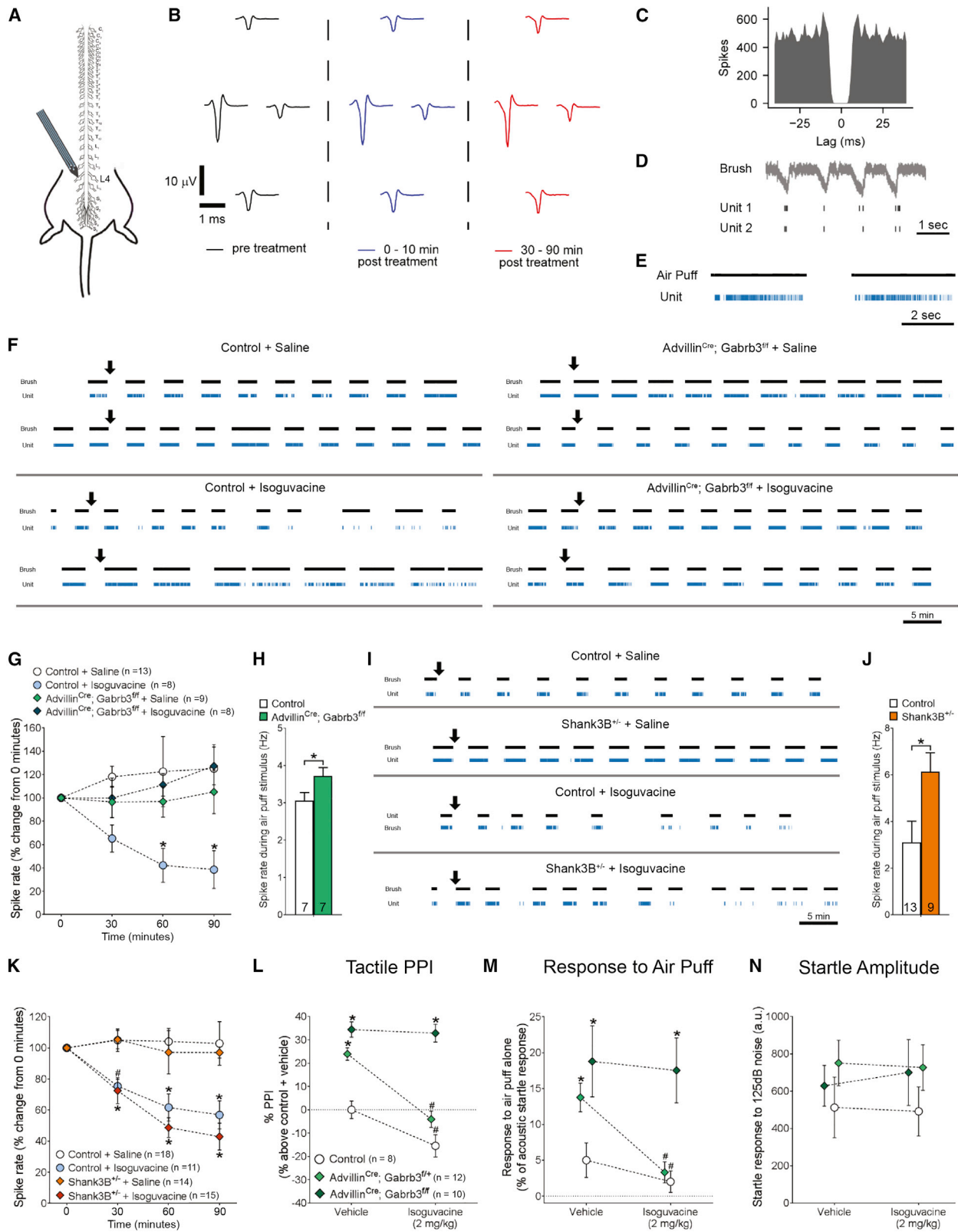
(M) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in control mice following i.p. administration of saline or isoguvacine at a range of concentrations. One-way ANOVA with post hoc Sidak's test (F[4, 113] = 8.005, *p* < 0.0001), asterisks above bars: **p* < 0.05 for comparisons between saline treatment and each isoguvacine concentration; asterisks above brackets: **p* < 0.05, for comparisons between indicated conditions.

(N) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in mice following i.p. administration of either saline or 2 mg/kg isoguvacine. Repeated-measures, two-way ANOVA with post hoc Sidak's test (F[7,277] = 16.44, *p* < 0.0001), **p* < 0.05 for comparisons between saline-treated mutants and saline-treated control littermates; #*p* < 0.05, for comparisons between saline and isoguvacine treatments within the same genotype.

(O) Response to a light air puff stimulus alone in mice following i.p. administration of either saline or 2 mg/kg isoguvacine treatment. Responses are expressed as percent of startle response to a 125 dB noise. Repeated-measures, two-way ANOVA with post hoc Sidak's test (F[7,277] = 9.650, *p* < 0.0001), **p* < 0.05 for comparisons between saline-treated mutants and saline-treated control littermates; #*p* < 0.05, for comparisons between saline and isoguvacine treatments within the same genotype.

(P) Magnitude of startle response to a 125 dB noise in mice following i.p. administration of either saline or 2 mg/kg isoguvacine treatment. Repeated-measures, two-way ANOVA, not significant.

Error bars indicate standard error of the mean. See also Figure S6 and Tables S5 and S7.



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site of action, isoguvacine did not cause sedation in any animal cohorts tested, as measured by startle amplitude (Figure 5P).

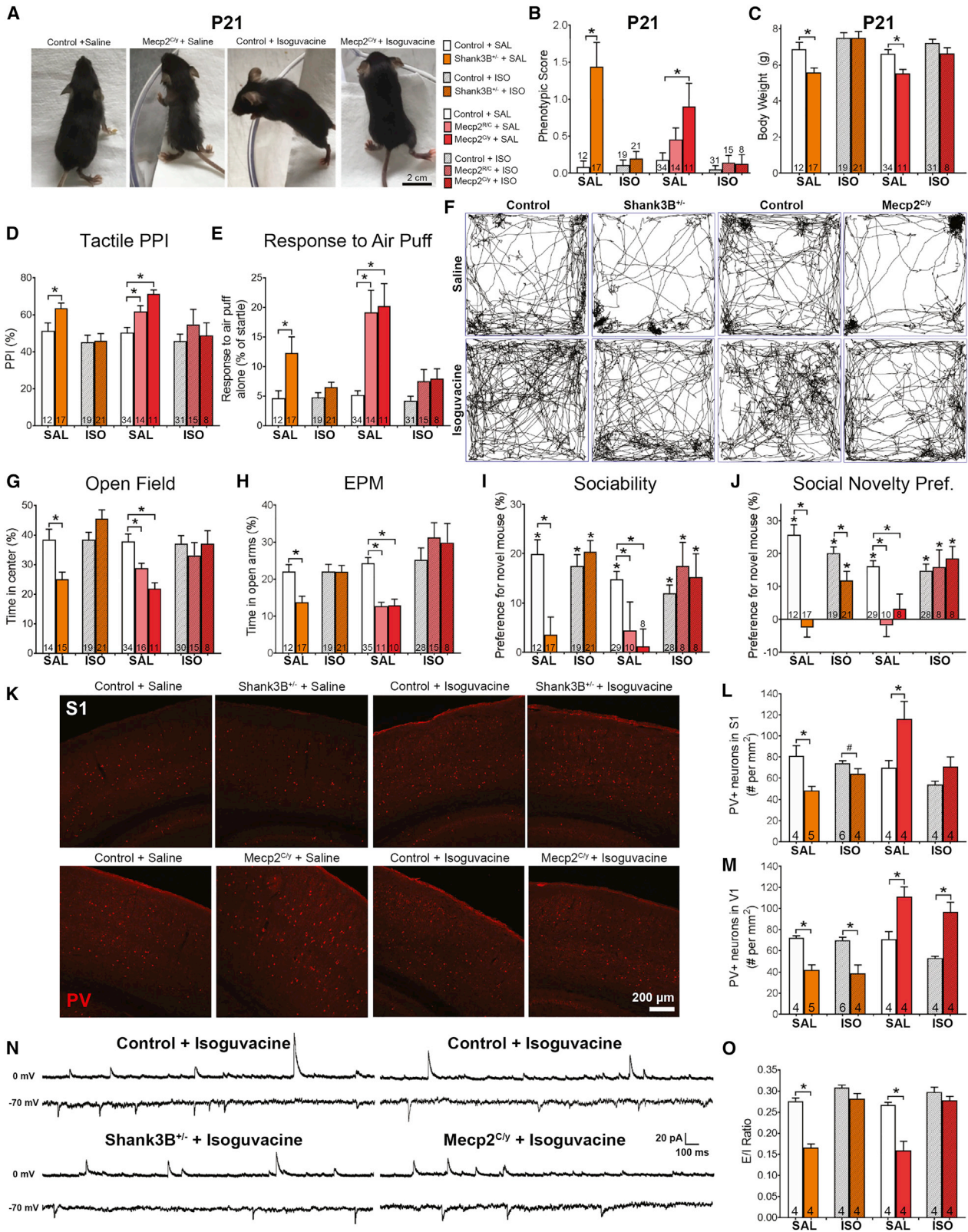
Isoguvacine Attenuates Tactile Sensitivity through Reduced Excitability of LTMRs

We next performed both *in vitro* and *in vivo* electrophysiological recordings to ask whether isoguvacine exerts its effects by acting directly on peripheral somatosensory neurons. Isoguvacine application reduced the hyperexcitability phenotype observed in large diameter neurons cultured from *Mecp2* and *Shank3* mutant mice, without affecting I_h (Figures S6A–S6G). We tested whether administration of isoguvacine would reduce mechanosensory neuron sensitivity *in vivo* by performing multi-unit electrode recordings in the L4 DRG of anesthetized mice (Figures 6A–6E; STAR Methods). Our findings indicate that while proprioceptor sensitivity and firing were unaffected, subcutaneous administration of isoguvacine increased response thresholds to light touch stimuli and reduced spiking in A β LTMRs *in vivo* (Figures 6F, 6G, S6P, and S6Q). These results are consistent with our finding that while LTMRs exhibit high expression levels of GABA_AR subunits, including GABRB3, proprioceptors do not (Figures 5J–5L and S6H). To test whether the effects of isoguvacine on cutaneous LTMR firing properties and tactile sensitivity are mediated through its direct activation of GABA_ARs in somatosensory neurons, we next performed a similar set of *in vivo* DRG recordings in mice with somatosensory neuron-specific, homozygous deletion of *Gabrb3* (*Advillin*^{Cre}; *Gabrb3*^{ff}) and control littermates (*Gabrb3*^{ff}). Because *Gabrb3* encodes the principal obligatory beta subunit of the GABA_AR in DRG neurons (Figures 5J–5L and S6H), somatosensory neurons in *Advillin*^{Cre}; *Gabrb3*^{ff} mutant mice are virtually devoid of

GABA_ARs (Orefice et al., 2016; Zimmerman et al., 2019). Sixty minutes following subcutaneous injection of isoguvacine (2 mg/kg), tactile stimulus-evoked (brush or air puff stimulus) LTMR responsiveness was decreased in control littermates, while no change in tactile-evoked LTMR responses were observed in control mice injected with saline (Figures 6F and 6G). Importantly, mice with somatosensory neuron-specific homozygous deletion of *Gabrb3* showed no changes in tactile-evoked LTMR spiking following administration of isoguvacine, indicating that isoguvacine exerts its effects by acting directly on GABRB3-containing GABA_ARs expressed on somatosensory neurons (Figures 6F–6H). We next asked whether isoguvacine treatment would attenuate LTMR sensitivity in a mouse model in which deficits in GABA_AR signaling is not the primary pathophysiological deficit. For this, *Shank3B*^{+/-} and control littermates were subjected to the same *in vivo* DRG recording preparation. LTMRs from saline-injected *Shank3B*^{+/-} mutant exhibited higher response rates to innocuous touch stimuli compared to LTMRs from control littermates injected with saline (Figures 6I and 6J), and isoguvacine attenuated firing in both *Shank3B*^{+/-} and control littermates (Figures 6I, 6K, and S6O). No differences in proprioceptor sensitivity were observed in any of the experiments (Figures S6P–S6R). Consistent with these electrophysiological measurements, mice with homozygous deletion of *Gabrb3* in peripheral sensory neurons (*Advillin*^{Cre}; *Gabrb3*^{ff} mice) exhibited no reduction in hairy skin sensitivity following isoguvacine treatment (Figures 6L and 6M). As above, startle amplitude was unaffected by isoguvacine (Figure 6N), indicating that isoguvacine was not sedating these mice. These findings indicate that a peripherally restricted GABA_AR agonist, isoguvacine, which attenuates tactile over-reactivity in several genetic and

Figure 6. Isoguvacine Attenuates Tactile Sensitivity through Reduced Excitability of Peripheral, Low-Threshold Mechanosensory Neurons

- (A) Diagram for *in vivo* dorsal root ganglion (DRG) multi-unit electrode recordings, showing tetrode placement into the left L4 ganglia.
- (B) Example single unit identified during the spike sorting process. Average waveform at each electrode site.
- (C) Example spiking autocorrelation for the unit depicted in 6B.
- (D) Activity traces of two putative single units in response to a brush stimulus.
- (E) Activity traces of a putative single unit in response to a light air puff stimulus (1 PSI).
- (F) Representative activity raster plots for multiple putative LTMRs in multiple mice over the duration of a recording experiment in controls and *Advillin*^{Cre}; *Gabrb3*^{ff} mice. Mice received a subcutaneous injection of either saline or isoguvacine (2 mg/kg) during the experiment, and activity of light-touch responsive units in response to a light brush stimulus was assessed over a 90-min period. Arrows indicate time of injection.
- (G) Relative firing frequency of LTMRs in response to a brush stimulus over the duration of each recording experiment, following subcutaneous injection of either saline or isoguvacine (2 mg/kg). Repeated-measures, two-way ANOVA with post hoc Dunnett's test ($F[3,136] = 9.326$, $p < 0.0001$), * $p < 0.05$.
- (H) Average baseline spike rate of LTMRs in response to an air puff stimulus (1 PSI), in control and *Advillin*^{Cre}; *Gabrb3*^{ff} mice. Student's t test, * $p = 0.049$.
- (I) Representative activity raster plots for putative LTMRs in multiple mice over the duration of recordings in controls and *Shank3B*^{+/-} mice. Mice received a subcutaneous injection of either saline or isoguvacine (2 mg/kg) during the experiment, and activity of light-touch responsive units was assessed over a 90-min period. Arrows indicate time of injection.
- (J) Average baseline spike rate of LTMRs in response to an air puff stimulus (1 PSI), in control and *Shank3B*^{+/-} mice. Student's t test, * $p = 0.0291$.
- (K) Relative firing frequency of LTMRs in response to a brush stimulus over the duration of each recording experiment, following subcutaneous injection of either saline or isoguvacine (2 mg/kg). Repeated-measures, two-way ANOVA with post hoc Dunnett's test ($F[3, 216] = 22.69$, $p < 0.0001$), * $p < 0.05$.
- (L) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in control, *Advillin*^{Cre}; *Gabrb3*^{ff} and *Advillin*^{Cre}; *Gabrb3*^{ff} mice following i.p. administration of 2 mg/kg isoguvacine (i.p., 2 mg/kg). Repeated-measures, two-way ANOVA with post hoc Sidak's test, * $p < 0.05$ for comparisons between mutant group to control littermates with saline; # $p < 0.05$, for comparisons between mutant condition with saline to same mutant group with isoguvacine.
- (M) Response to a light air puff stimulus alone in mice following i.p. administration of either saline or 2 mg/kg isoguvacine treatment. Responses are expressed as percent of startle response to a 125 dB noise. Repeated-measures, two-way ANOVA with post hoc Sidak's test, * $p < 0.05$ for comparisons between mutant group to control littermates with saline; # $p < 0.05$, for comparisons between mutant condition with saline to same mutant group with isoguvacine.
- (N) Magnitude of startle response to a 125 dB noise in mice following i.p. administration of either saline or 2 mg/kg isoguvacine treatment. Repeated-measures, two-way ANOVA with post hoc Sidak's test, not significant.
- Error bars indicate standard error of the mean. See also Figure S6 and Table S7.



(legend on next page)

environmental ASD mouse models, exerts its effects through direct activation of GABA_AR expressed on LTMRs.

Chronic Treatment with a Peripherally Restricted GABA_AR Agonist Improves Some ASD Phenotypes in Two Mouse Models for ASD

Our results raised the possibility that peripherally restricted GABA_AR agonists may be useful for treating tactile over-reactivity, and potentially anxiety-like behaviors and certain social impairments in different ASD mouse models, if treatment is administered during early postnatal development. Therefore, we tested whether chronic treatment of *Mecp2* and *Shank3* germline mutant mice with isoguvacine beginning shortly after birth improves any of the ASD-related phenotypes commonly observed in adulthood. Following a 6-week treatment regimen, isoguvacine was detected in the liver, but not brain or SC (Figure S7A). When isoguvacine-treated mice were first compared to saline-treated groups at weaning age (P21), we noticed an improvement in their overall body health or phenotypic score (Figures 7A and 7B; STAR Methods). Moreover, while saline-treated *Shank3B* and *Mecp2* mutant mice displayed reduced bodyweight at P21 compared to control littermates, isoguvacine-treated mutant mice were not different in weight from control mice (Figure 7C). Chronic isoguvacine administration also improved multiple ASD-associated behavioral phenotypes, including tactile over-reactivity, anxiety-like behaviors, social impairments, as well as PV⁺ neuron abnormalities in S1 and BLA, and normalized E/I ratios in S1 of 8-week-old mice (Figures 7D–7O, S7I–S7K, and S7N–S7S; Table S6). Chronic isoguvacine administration, however, did not improve texture discrimination deficits, increased acoustic PPI performance, memory impairments, PV⁺ neuron abnormalities in V1, or altered E/I ratios in V1 in either *Shank3* or *Mecp2* mutants (Figures S7C, S7E, S7H, and S7T–S7V). Motor impairments were also not improved in *Mecp2* mutants, nor was the overgrooming phenotype rescued in *Shank3* mutants (Figures S7D, S7L, and S7M). Taken together, treatment with a peripherally restricted

GABA_AR agonist during early postnatal development may provide a therapeutic strategy for improving tactile over-reactivity and a subset of other key features of ASD, including anxiety, without causing sedation and other undesirable effects of activating brain GABA_AR during development.

DISCUSSION

Aberrant sensory reactivity is now regarded as a diagnostic feature of ASDs. The present study adds to a growing body of work demonstrating that somatosensory neurons are dysfunctional and contribute to behavioral phenotypes in a range of genetic and environmental models for ASD, including *Mecp2*, *Gabrb3*, *Shank3*, *Cntnap2*, *Fmr1*, and MIA mouse models (Bhattacharjee et al., 2017; Chen et al., 2014; Dawes et al., 2018; Han et al., 2016; Oginsky et al., 2017; Orefice et al., 2016; Perche et al., 2018; Price and Melemedjian, 2012; present study). A main finding of the present work is that tactile over-reactivity in ASD models can arise from distinct cell-autonomous, pathophysiological mechanisms. Tactile over-reactivity may result from loss of GABA_AR signaling and PSI in somatosensory neurons, as is the case for *Mecp2* and *Gabrb3* mutants, or loss of potassium channel function leading to somatosensory neuron hyper-excitability, as seen here in *Shank3* mutants. It is noteworthy that mutations in *Mecp2* and *Shank3* may differentially affect sensitivity of LTMRs and small diameter nociceptive neurons, which may explain the seemingly paradoxical findings that at least some ASD patients can exhibit both hypersensitivity and aversion to light touch but also decreased responsiveness to noxious stimuli (Downs et al., 2010; Tomchek and Dunn, 2007). Consistent with this, reduced sensitivity to painful thermal and chemical stimuli is observed in mice with conditional deletion of *Shank3* in sensory neurons (Han et al., 2016).

A second principle to emerge from the present work is that loss of either *Mecp2* or *Shank3* in peripheral sensory neurons leads to changes in neurochemical and functional properties of

Figure 7. Chronic Administration of Isoguvacine Improves Tactile Over-Reactivity, Region-Selective Cortical Abnormalities, and Some ASD-Related Behaviors in *Mecp2* and *Shank3* Mutant Mice

- (A) Representative images of P21 control and *Mecp2*^{C/y} mutant mice treated daily from P1–21 with either saline or isoguvacine (2 mg/kg).
- (B) Average phenotypic score of P21 *Shank3B*^{+/-} or *Mecp2*^{C/y} mutant mice and control littermates treated daily with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (C) Average bodyweight of P21 *Shank3B*^{+/-} or *Mecp2*^{C/y} mutant mice and control littermates treated daily with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (D) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in *Shank3B*^{+/-}, *Mecp2*^{R/C}, or *Mecp2*^{C/y} mutant mice and control littermates treated daily from P1–P42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (E) Response to a light air puff stimulus alone in *Shank3B*^{+/-}, *Mecp2*^{R/C}, or *Mecp2*^{C/y} mutant mice and control littermates treated daily from P1–P42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (F) Representative activity traces in the OF test. OF chamber is 40 cm by 40 cm.
- (G) Percent time spent in the center of the OF chamber. One-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (H) Percent time spent in the open arms of the EPM. One-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (I and J) Preference index for the percentage of time spent investigating the novel mouse in the "Sociability" (I) or "Social Novelty Preference" (J) portion of the 3-chamber social interaction test. One-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (K) Representative IHC images of coronal S1 brain sections showing PV immunoreactivity in *Shank3B*^{+/-} or *Mecp2*^{C/y} mutant mice and control littermates treated daily from P1–P42 with either saline or isoguvacine (2 mg/kg).
- (L and M) Quantification of the number of PV-positive (PV⁺) neurons in S1 (L) and V1 (M). One-way ANOVA with post hoc Tukey's test, *p < 0.05; #p < 0.10.
- (N) Representative traces showing sEPSCs (–70 mV hold) and sIPSCs (0 mV hold) from S1 slices from *Shank3B*^{+/-} or *Mecp2*^{C/y} mutant mice and control littermates treated daily from P1–P42 with either saline or isoguvacine (2 mg/kg).
- (O) Quantification of excitatory/inhibitory (E/I) ratio in S1 slices from control and mutant mice. One-way ANOVA with post hoc Sidak's test, *p < 0.05. Error bars indicate standard error of the mean. See also Figure S7 and Tables S6 and S7.

brain circuits. We found that PV⁺ interneurons in S1 and BLA are adversely affected in mice lacking *Mecp2* or *Shank3* in somatosensory neurons (Table S7). We speculate that alterations in the number of PV⁺ neurons or PV expression in cortex reflect homeostatic mechanisms for increasing inhibitory neuron response rates under conditions of enhanced sensory drive to the cortex, and these alterations differ depending on the developmental time point at which sensory drive is altered. This is consistent with prior findings that both increased PV⁺ neuron number and decreased PV expression are associated with enhanced inhibitory transmission and reduced cortical response rates to sensory stimuli (Volman et al., 2011; Vreugdenhil et al., 2003) as well as recent findings that changes in sensory cortex E/I balance observed in ASD models may reflect adaptations to altered sensory input from the periphery (Antoine et al., 2019). It is important to note, however, that while peripheral somatosensory neuron dysfunction contributes to altered cortical circuit development, loss of ASD-related genes within the brain is also likely to contribute to altered sensory cortex microcircuit function in ASD models. While our findings implicate *Mecp2* and *Shank3* function in peripheral sensory neurons for normal tactile, anxiety-like behaviors, and some social behaviors, we also emphasize that loss of *Mecp2* or *Shank3* in peripheral sensory neurons does not recapitulate all ASD behavioral phenotypes observed in the germline mutation models. This includes memory impairments, motor deficits, respiratory function and early lethality in *Mecp2* mutants, and memory impairments and over-grooming behaviors observed in *Shank3* mutants.

A third general finding of the present work is the extent to which sensory neuron dysfunction contributes to aberrant behavior in disparate ASD models varies greatly with respect to the timing or developmental onset of sensory dysfunction. While early developmental restoration of either *Mecp2* or *Shank3* function in sensory neurons improves hairy skin sensitivity, some aspects of social behavior, and anxiety-like behaviors, P28 restoration improves hairy skin sensitivity and sociability but has no effect on social novelty recognition preference and anxiety-like behaviors. This is consistent with prior studies in which global restoration of *Shank3* at P21 was sufficient to normalize sociability behaviors but not anxiety-like behaviors (Mei et al., 2016; Wang et al., 2017).

Our findings lead us to propose a pharmacological approach to augment GABA_A receptor signaling in LTMRs during early postnatal development as a novel therapy for ASD. The logic behind the “LTMR GABA_AR hypothesis” is based on the following observations: (1) ASD gene dysfunction in mechanosensory neurons causes altered physiological properties of LTMRs, including hypersensitivity and reduced PSI in the SC, which contribute to tactile over-reactivity in ASD mouse models (Orefice et al., 2016; present study); (2) GABA_ARs are present all along myelinated axons of peripheral nerves (Zeilhofer et al., 2012), and peripheral release of GABA controls somatosensory neuron sensitivity (Carlton et al., 1999; Hanack et al., 2015; Obradovic et al., 2015); (3) the ASD-associated gene *Gabrb3*, which encodes the principal obligatory beta subunit of GABA_ARs in DRG neurons in both mice (Orefice et al., 2016; Zheng et al., 2019; Zimmerman et al., 2019; present study) and humans (Flegel et al., 2015; Ray et al., 2018), functions cell autonomously in LTMRs to control

tactile sensitivity (Orefice et al., 2016; present study); (4) GABA acts directly on DRG neurons to reduce excitability *in vitro* (Du et al., 2017; present study); (5) administration of the peripherally restricted GABA analog, isoguvacine, attenuates LTMR firing properties and tactile sensitivity *in vivo* in a manner that is dependent on GABA_ARs present on LTMRs (present study); (6) isoguvacine normalizes tactile over-reactivity in five genetic and one environmental model for ASD (present study); and (7) chronic treatment with isoguvacine beginning at early postnatal ages in two distinct models for ASD improves overall body condition, body weight, PV⁺ interneuron alterations in S1 and BLA, E/I ratios in S1, as well as anxiety-like behaviors and some social impairments in young adult mice (present study). In line with the LTMR GABA_AR hypothesis, we speculate that other new treatment strategies that show promise for ASD may also work through affecting peripheral nerve function, including bumetanide, which is largely peripherally restricted (He et al., 2018; Römermann et al., 2017). We propose that a benefit of peripheral restriction is to avoid complications associated with direct brain actions and adverse effects on brain development. Consistent with this, peripheral administration of isoguvacine failed to penetrate the brain to an appreciable extent and did not lead to sedation, as compared to brain-penetrating GABA_AR PAMs. Thus, we propose that GABA_AR agonists, GABA reuptake inhibitors, or GABA_AR PAMs that are peripherally restricted may reduce tactile over-reactivity and improve brain microcircuit function and related ASD behaviors observed in certain patients with ASD, while minimizing or avoiding entirely potentially detrimental effects on brain development observed in clinical use of classical, FDA-approved GABA_A drugs (Kodish et al., 2011), all of which penetrate the BBB (Groeneveld et al., 2016). Peripherally restricted methods for augmenting GABA_AR signaling may also have applicability in other diseases and disorders in which touch over-reactivity is present, such as mechanical allodynia in neuropathic pain states, sensory processing disorder, and schizophrenia. Whether acute treatment of ASD patients with peripherally restricted compounds to augment GABA_AR signaling normalizes tactile over-reactivity, and whether chronic treatment beginning at early ages ameliorates some ASD-associated behaviors, awaits the development of safe, peripherally restricted compounds and their testing in patient trials.

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

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● QUANTIFICATION AND STATISTICAL ANALYSIS

SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at <https://doi.org/10.1016/j.cell.2019.07.024>.

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AUTHOR CONTRIBUTIONS

L.L.O. and D.D.G. conceived the study. L.L.O. performed electrophysiology experiments. L.L.O. executed behavioral and histological experiments with assistance of J.R.M., D.T.M., A.T., and S.M.M. M.F.W. and G.F. generated the *Shank3*^{flxed} mouse. M.Y. made the GABRB3-expressing virus. L.L.O. and R.M.F. performed the bio-distribution experiments with the help of J.R.M. and S.M.M. A.M.C., A.J.E., and G.R. developed the *in vivo* DRG MEA recording preparation. L.L.O. and D.D.G. wrote the paper, with input from all authors.

DECLARATION OF INTERESTS

L.L.O. and D.D.G. have patents pending on the use of peripherally restricted GABA_A receptor drugs for the treatment of tactile over-reactivity in ASD and other disorders.

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STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
Goat anti GABRB3	Life Technologies	PA5-19060; RRID:AB_10985298
Mouse anti GAD67	Fisher Scientific	MAB5406; RRID:AB_2278725
Rabbit anti HCN1	Alomone Labs	APC-056; RRID:AB_2039900
Alexa 647-conjugated IB4	Life Technologies	I16520-100
Rabbit anti MeCP2	Michael Greenberg	n/a
Goat anti PARVALBUMIN	Swant	PVG-213; RRID:AB_2650496
Rabbit anti PARVALBUMIN	Swant	PV-27; RRID:AB_2631173
Rabbit anti dsRED	Clontech	632496; RRID:AB_10013483
Mouse anti Shank3	Abcam	Ab193307; RRID:AB_2687405
Guinea pig anti vGLUT1	Millipore	AB5905; RRID:AB_2301751
Bacterial and Virus Strains		
AAV2.9 hSyn-FLEX	Addgene	#84481; RRID:Addgene_84481
Chemicals, Peptides, and Recombinant Proteins		
Dexamethasone	Henry Schein	Cat. # 2459
Isoguvacine	Sigma Aldrich	Cat. #G002
Isoflurane (Isothesia)	Henry Schein	Cat. # 029405
Midazolam	Henry Schein	Cat. # 054788
Nipecotnic Acid	Sigma Aldrich	Cat. # 211672
Polyinosinic: polycytidylic acid	Millipore	Cat. # 528906
Tamoxifen	Toronto Research Chemicals	Cat. # 10540-29-1
Urethane	Sigma Aldrich	Cat. # U2500
Critical Commercial Assays		
IonFlux HTTM Automated Patch Clamp System	Eurofins	N/A
In-Fusion HD Cloning Plus Kit	Takara	638911
Experimental Models: Organisms/Strains		
Advillin ^{Cre}	Fan Wang; RRID:IMSR_JAX:032536	Hasegawa et al., 2007
16p11.2 df null	Jackson Laboratory	RRID:IMSR_JAX:013128
Advillin ^{CreERT2}	John Wood; RRID:IMSR_JAX:032027	Lau et al., 2011
Cdx2 ^{Cre}	Eric Fearon; RRID:IMSR_JAX:009350	Akyol et al., 2008
Cntnap2 null	Jackson Laboratory	RRID:IMSR_JAX:028635
Fmr1 null	Jackson Laboratory	RRID:IMSR_JAX:003025
Gabrb3 ^{floxed}	Jackson Laboratory	RRID:IMSR_JAX:008310
Mecp2 ^{floxed}	Jackson Laboratory	RRID:IMSR_JAX:006847
Mecp2 ^{R306C}	Michael Greenberg	Lyst et al., 2013
Mecp2 ^{STOP}	Jackson Laboratory	RRID:IMSR_JAX:006849
Shank3B Null	Jackson Laboratory	RRID:IMSR_JAX:017688
Shank3 ^{Δex4-9} null	Jackson Laboratory	RRID:IMSR_JAX:017890
Shank3 ^{floxed}	This paper	N/A
Shank3 ^{FX}	Guoping Feng; RRID:IMSR_JAX:028800	Mei et al., 2016
Oligonucleotides		
Primers for mouse genotyping (see Table S8)	This paper	N/A
Recombinant DNA		
AAV.hSYN.FLEX.GABRB3.t2A.mCHERRY	This paper	N/A

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REAGENT or RESOURCE	SOURCE	IDENTIFIER
Software and Algorithms		
ImageJ	Schneider et al., 2012	https://imagej.nih.gov/ij/
JRClust	Jun et al., 2017	https://github.com/JaneliaSciComp/JRCLUST

LEAD CONTACT AND MATERIALS AVAILABILITY

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, David Ginty (david_ginty@hms.harvard.edu).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

All procedures performed in this study were approved by the Harvard Medical School Institutional Animal Care and Use Committee (IACUC). Male and female mice of mixed genetic backgrounds (C57BL/6J, 129/SvEv, CD1) were used for these studies. The only exceptions were germline mutant mice (*Mecp2*^{R306C}, *Shank3B*^{+/-} and *Shank3*^{Δex4-9}), which were backcrossed for at least 5 generations to a C57BL/6J background. Testing was done beginning at 6 weeks of age, and in most cases, behavioral testing was completed by 8 weeks of age. All behavioral testing performed in female mice was completed prior to the start of estrous cycles. The ages of mice used for histological and electrophysiological experiments are denoted in the appropriate [Method Details](#) and relevant Figure Legends. Mice were weaned and ear notched for genotyping at P21 (+/- 2 days). All animals were group housed, with control and mutant animals in the same litters and cages. Littermates from the same genetic crosses were used as controls for each group, to control for variability in mouse strains/backgrounds. No differences were observed between wild-type animals and any single floxed or Cre control groups for any of the tests performed. Male and female mice of the same genotype were first analyzed separately to assess potential sex-related differences in behaviors. If no differences were observed, male and female mice of the same genotype were grouped together for final analyses. Behavioral and histological differences were found between hemizygous null male *Mecp2*^{C/y} and heterozygous female *Mecp2*^{R/C} mice. For this reason, these two groups were analyzed separately and compared to male and female littermate controls.

Generation of Shank3 conditional knockout mouse

The targeting vector flanked Exons 13-16 (PDZ domain) of the Shank3 gene with loxP sites and a NEO cassette. Chimeric mice were crossed to C57BL/6J females from Jackson Labs. Germline transmission was assessed through genotyping PCR of mouse tail DNA, using primers pFW (Sh3cKO) Gen 1a and pRV (Sh3cKO) Gen 3b for the wild-type allele (positive band = 188 base pairs) and the floxed allele (positive band = 228 base pairs). The F1 hybrids were crossed to C57BL/6J β-Actin Flp mice to excise the NEO cassette. The floxed mice were then backcrossed to C57BL/6J mice for 5 generations. After the 5th generation, speed congenic genotyping PCRs were conducted to determine the approximate purity of the background. Only mice showing > 95% C57BL/6J background were used for subsequent matings. Backcrossed Shank3floxed mice were then bred with C57BL/6J β-Actin Cre mice to produce germline knockouts of the floxed allele (termed *Shank3*^{flxed} mice). Genotypes were determined by PCRs using the pFW (Sh3cKO) Gen 1a and pRV (Sh3cKO) Gen 3b for the knockout allele (positive band = 290 base pairs). pFW (Sh3cKO) Gen 1a forward primer: 5'- CAG CAT TTA TAC CTG ACT GTG AAG C -3'; reverse primer: 5'- GGG AGT AGA GCT CAG ATA ACC -3'.

Mouse Lines and Genotyping

Mice were group housed with littermates in standard housing on a 12-hour light/dark cycle. Tail biopsies were taken at weaning (P21, +/- 2 days), which were used for genetic identification.

Shank3B null mice were obtained from the Jackson Laboratory (017688) and were previously described ([Peça et al., 2011](#)). The following primers were used to identify the null allele: common forward 5'- GAG ACT GAT CAG CGC AGT TG -3'; wild-type reverse 5'- TGA CAT AAT CGC TGG CAA AG -3'; mutant reverse 5'- GCT ATA CGA AGT TAT GTC GAC TAG G -3'.

Shank3^{Δex4-9} null mice were obtained from the Jackson Laboratory (017890) and were previously described ([Bozdagi et al., 2010](#)). The following primers were used to identify the null allele: common forward 5'- TGG GAT GTG AGA GTG ACC AG-3'; wild-type reverse 5'- AGG AGG TCA GTG GCG TTG T -3'; mutant reverse 5'- AAT TCT GTT CAG TCC ACA CAG G -3'.

Fmr1 null mice were obtained from the Jackson Laboratory (003025) and were previously described ([Bakker et al., 1994](#)). The following primers were used to identify the null allele: mutant forward 5'- CAC GAG ACT AGT GAG ACG TG -3'; wild-type forward 5'- TGT GAT AGA ATA TGC AGC ATG TGA-3'; common reverse 5'- CTT CTG GCA CCT CCA GCT T -3'.

16p11.2 df null mice (*16p11.2*) were obtained from the Jackson Laboratory (013128) and were previously described ([Horev et al., 2011](#)). The following primers were used to identify the null allele: mutant forward 5'- ACC TGG TAT TGA ATG CTT GC -3'; wild-type forward 5'- CCT GAG CCT CTT GAG TGT CC -3'; mutant reverse 5'- TGG TAT CTC CAT AAG ACA GAA TGC -3'; wild-type reverse 5'- GTC GGT TCA GGT GGT AGA CG -3'.

Hcn1 null mice were obtained from the Jackson Laboratory (028301) and were previously described (Nolan et al., 2003). The following primers were used to identify the null allele: mutant forward 5'-AGA GAA ATC ATT CCC CGT GA-3'; wild-type forward 5'-CAC CTG CTA CGC AAT GTT TG -3'; common reverse 5'-ATT GGG CAC TAC ACG CTA GG -3'.

Shank3^{FX} mice were previously described (Mei et al., 2016). The following primers were used to identify the mutant allele: for the wild-type allele, forward 5'-CGT TTG ACA CAC ATA AGC ACC -3' and reverse 5'-CTC CAC CTA GCT GAA TTT CCC -3' were used to produce a band of 340 bp. For the knockout (FX) allele, forward 5'-CGT TTG ACA CAC ATA AGC ACC-3' and reverse 5'-GCT GAC ATC ACA TTG CTG CC -3' were used to produce a band of 481 bp. For the rescue allele, forward 5'-CGT TTG ACA CAC ATA AGC ACC -3' and reverse 5'-CTC CAC CTA GCT GAA TTT CCC -3' were used to produce a band of 408 bp.

Mecp2^{R306C} mice were obtained from Michael Greenberg and were previously described (Lyst et al., 2013). The following primers were used to identify the mutant allele: forward 5'-GGA TTG TGG AAA AGC CAG-3'; and reverse 5'-ATG ACC TGG GCA GAT GTG GTA G-3'.

Mecp2 floxed mice were obtained from the Jackson Laboratory (006847). The floxed *Mecp2* sequence was identified using the following primers: forward 5'-TGG TAA AGA CCC ATG TGA CCC AAG-3' and reverse 5'-GGC TTG CCA CAT GAC AAG AC-3'. The following primers were used to identify the null allele (post Cre excision), as well as to check for off-target Cre expression in *Advillin^{Cre}* mice: forward 5'-TGG TAA AGA CCC ATG TGA CCC AAG-3'; and post-Cre excision reverse 5'-TCC ACC TAG CCT GCC TGT ACT TTG-3'.

Mecp2^{STOP} mice were obtained from the Jackson Laboratory (006849) and were previously described (Guy et al., 2007). The following primers were used to identify the mutant allele: forward common 5'-AAC AGT GCC AGC TGC TCT TC-3'; wild-type reverse 5'-CTG TAT CCT TGG GTC AAG CTG -3' and mutant reverse 5'-GCC AGA GGC CAC TTG TGT AG -3'.

Gabrb3 floxed mice were obtained from the Jackson Laboratory (008310) and were previously described (Ferguson et al., 2007). The floxed *Gabrb3* sequence was identified using the following primers: forward 5'-ATT CGC CTG AGA CCC GAC T -3' and reverse 5'-GTT CAT CCC CAC GCA GAC -3'.

Cntnap2 null mice were obtained from the Jackson Laboratory (028635) and were previously described (Gordon et al., 2016). The mutant *Cntnap2* sequence was identified using the following primers: wild-type forward 5'-TGC CCT CCT AGA AAG TAA ATG C T-3'; mutant forward 5'-GCC AGA GGC CAC TTG TGT AG -3' and common reverse 5'-TCC TCT CTT CAT GCA CAC TAT GA -3'.

Advillin^{Cre} mice were obtained from Fan Wang (Duke University) and were previously described (Hasegawa et al., 2007). The *Advillin^{Cre}* transgene was identified using the following primers: 5'-CCC TGT TCA CTG TGA GTA GG -3'; reverse 5'-AGT ATC TGG TAG GTG CTT CCA G -3'; and internal control 5'-GCG ATC CCT GAA CAT GTC CAT C -3'.

Advillin^{CreERT2} mice were obtained from John Wood and previously described (Lau et al., 2011). The *Advillin^{CreERT2}* transgene was identified using the following primers: 5'-CCC TGT TCA CTG TGA GTA GG -3'; 5'-AGT ATC TGG TAG GTG CTT CCA G -3'; and 5'-GCG ATC CCT GAA CAT GTC CAT C -3'.

Cdx2^{Cre} mice were obtained from Eric Fearon, and previously described (Akyol et al., 2008). The *Cdx2^{Cre}* transgene was identified using the following primers: forward 5'-CTC GAC GTC TCC AAC CAT TG -3'; and reverse 5'-ATC TTC AGG TTC TGC GGG AA -3'.

Proper expression of each floxed allele using each Cre transgene was assessed using PCR and histological verification. *Advillin^{Cre}* or *Advillin^{CreERT2}* animals with post-Cre excision expression in tail biopsy tissue or animals with transgene expression and recombination in the spinal cord or brain were excluded from analyses.

METHOD DETAILS

Generation and use of AAV.hSYN.FLEX.GABRB3.t2A.mCHERRY

AAV cloning and production: cDNA of mouse *Gabrb3* was purchased from GE Healthcare. *Gabrb3* cDNA and T2A-mCherry were cloned into an AAV2.9 plasmid that contains the hSyn-FLEX cassette (Addgene #84481) by In-Fusion cloning kit (Takara). AAV particles were produced by The Boston Children's Hospital Viral Core, with a titer of 8E+12 vg/mL. 2 μ L of AAV.hSYN.FLEX.GABRB3.t2A.mCHERRY ('AAV.FLEX.GABRB3.mCHERRY') were diluted into 18 μ L of saline, and then injected i.p. into each pup at both P5 and P6. Pups were returned to their home cage each day following injections. Mice harboring an arginine-to-cysteine missense mutation in *Mecp2* (*Mecp2^{R306C}*), which is a common allele found in patients with Rett Syndrome (RTT) (Lyst et al., 2013), were used in these studies because *Mecp2^{R306C}* mutant mice live longer than mice harboring the *Mecp2* null allele, allowing a full two-week behavioral assessment at six to eight weeks of age prior to the development of severe breathing abnormalities. *Mecp2^{R306C}* littermate mice lacking the *Advillin^{Cre}* allele and injected with AAV.FLEX.GABRB3.mCHERRY served as a control, as did wild-type littermates with and without the *Advillin^{Cre}* allele. We found that i.p. injection of *Advillin^{Cre}*; *Mecp2^{R306C}* mice, but not *Mecp2^{R306C}* mice lacking the *Advillin^{Cre}* allele, with AAV.FLEX.GABRB3.mCHERRY led to transduction of all types of DRG sensory neurons, a large fraction of which are LTMRs, and a dramatic increase in the amount of GABRB3 puncta at presynaptic terminals of A β -LTMRs and A δ -LTMRs in the spinal cord (SC) dorsal horn (Figures 4A-4C, S5A, and S5B).

Tamoxifen Administration

Tamoxifen was administered to *Advillin^{CreERT2}* mice to allow for the excision of floxed alleles (either *Shank3^f*, *Shank3^{FX}*, *Mecp2^f* or *Mecp2^{STOP}*) in peripheral somatosensory neurons of adult mice. Intraperitoneal injections of tamoxifen (1 mg per day, Toronto Research Chemicals) were administered to mice for 5 consecutive days, from either P5-9, P10-14 or P28-32. All mice in this study

shown for *Advillin*^{CreERT2} mouse lines, including *Advillin*^{CreERT2} and floxed allele controls in these groups, received this tamoxifen regimen, and no changes in health or behaviors were observed in either sets of controls compared to non-tamoxifen treated animals. A five-day tamoxifen dosing regimen resulted in recombination of target genes in > 90% of DRG neurons, while *Shank3* and *Mecp2* gene expression was unaltered in the brains or spinal cord of these mice (Figures S4C–S4E).

Poly I:C Administration

Administration of polyinosinic: polycytidylic acid (Poly I:C) was performed as previously described (Shin Yim et al., 2017). Timed-pregnant female mice were ordered from Charles River, and were verified to be positive for segmented filamentous bacteria. E12.5 pregnant female mice were weighed and injected with a single dose of poly(I:C) (20mg/kg i.p., Millipore) or saline as a vehicle control. Each dam was returned to its cage and left undisturbed until the birth of its litter.

Acute Drug Treatment to Assess Tactile Sensitivity

Mutant mice and their control littermates were subjected to two consecutive days of tactile PPI testing (see below for behavioral testing). Animals received i.p. injection of either vehicle or drug (midazolam 2mg/kg or isoguvacine 2mg/kg). After a mouse was injected, it was returned to their home cage for 30 minutes until they were subjected to tactile PPI. Litters were counterbalanced for treatment, such that half of the animals received saline on the first day and the other half of the litter received drug treatment on the first day. The following day, treatment regimens were reversed, and the mice were subjected to a second round of tactile PPI testing. For mutant animals of each line, percent PPI data are represented as percent of control littermate performance for saline treatment.

Chronic Isoguvacine Administration

Mecp2^{R306C} and *Shank3B*^{+/-} mice were chosen for these analyses because both are well-established models for ASD that exhibit behavioral abnormalities with high penetrance and severity. *Shank3B*^{+/-} or *Mecp2*^{R/C} female mice were bred to male c57Bl6/J mice. Once females were visibly pregnant (E17–20), they were singly-housed. On the day after each litter was born (P1), pups were weighed and administered i.p. injection of either isoguvacine (2mg/kg) or saline control. Pups were weighed and injected every day thereafter with the same treatment, ending on P42. At P21, pups were weighed, photographed, assessed for overall phenotype, and weaned. Females were counterbalanced such that half of the females had litters that received isoguvacine first, while the other half of the females' litters received saline treatment first. Whole litters received the same treatment regimen. Once a litter was weaned, mothers were re-mated with the same c57Bl6/J male mouse, and the treatment condition of the subsequent litter was reversed. At P42, mice began behavioral testing.

Behavioral Testing

Male and female mice of mixed genetic backgrounds (C57BL/6J, 129/SvEv, CD1) were used for behavioral analyses. The only exceptions were germline mutant mice (*Mecp2*^{R306C}, *Shank3B*^{+/-} and *Shank3*^{Δex4-9}), which were backcrossed for at least 5 generations to a C57BL/6J background. Testing was done beginning at 6 weeks of age, and in most cases, behavioral testing was completed by 8 weeks of age. All behavioral testing performed in female mice was completed prior to the start of estrous cycles. Mice were weaned and ear notched for genotyping at P21 (+/- 2 days). All animals were group housed, with control and mutant animals in the same litters and cages. Littermates from the same genetic crosses were used as controls for each group, to control for variability in mouse strains/backgrounds. No differences were observed between wild-type animals and any single floxed or Cre control groups for any of the tests performed. Male and female mice of the same genotype were first analyzed separately to assess potential sex-related differences in behaviors. If no differences were observed, male and female mice of the same genotype were grouped together for final analyses. Prior to behavioral testing, animals were brought into the procedure room and allowed to habituate to the room for 30 minutes. Animal numbers per group for behavioral tests are indicated in figures and/or supplemental tables. All behavioral analyses were performed and analyzed by experimenters who were blinded to genotype and treatment.

Ear Notching

Ear notching was employed as the method of identification for all animals used in behavioral experiments, as toe tagging or metal ear tags can affect animals' ambulatory behavior. Ear notching was performed using an ear punch device (Kent Scientific) on mice at weaning age (3 weeks of age). For this, mice were restrained by the scruff and the ear punch was used to create holes and/or notches in the ears, following the "Universal Mouse Numbering System." This method of identification allowed experimenters to identify mice without unnecessary handling of the mouse.

Whisker Plucking

To avoid the confound of whisker sensing by mice during NORT experiments, bilateral removal of all mystacial pad vibrissae was performed 3 days prior to the start of experiments. Mice were placed under isoflurane anesthesia and all mystacial vibrissae were plucked with tweezers. Whisker removal caused a small but significant increase in grooming time during a 20-minute open field test, but no differences in speed of movement, ambulatory time or jump time were observed following whisker removal (Orefice et al., 2016). Animals with and without whiskers also performed comparably on the textured NORT test, but removal of whiskers

did promote object investigation using the glabrous skin on paws over whisking/nose poke investigations (Orefice et al., 2016). Whisker removal also caused no changes in anxiety-like behavior, as measured by time spent in the open arms of an elevated plus maze (Orefice et al., 2016). Whiskers begin to regrow 8-12 days after plucking. NORT was completed in the first week following whisker removal, and therefore whiskers were only plucked once. Whiskers were allowed to regrow thereafter, and animals had regrown whiskers by the time social behavior tests were performed.

Open Field Test

On the first day of behavioral testing, animals were subjected to the open field test. During this test, an animal was placed in an empty plexiglass arena (40 cm x 40 cm x 40 cm) and allowed to explore for 10 minutes, under dim lighting. The outside walls of the chamber were opaque. The position of the mouse was tracked using custom MATLAB scripts. A portion of videos were blindly hand-scored to verify accuracy. Distance traveled and time spent away from walls (time in center) were analyzed per 10-minute video.

Novel Object Recognition Test (NORT)

NORT testing was performed as previously described (Orefice et al., 2016). NORT was performed in the same plexiglass arena used for open field testing. The cube-shaped objects were designed to promote climbing, which enhances novel object recognition performance requiring texture discrimination via glabrous skin on the paws (Heyser and Chemero, 2012). In this test, mice were first habituated to an open field chamber by allowing free exploration of an empty chamber for 10 minutes during each of two consecutive days (Figure 1D). The first day of habituation was used as the open field test (see above for details).

Each of the three subsequent testing days included two sessions. In the first session (learning phase), the mouse was placed in the testing arena, equidistant to, and facing away from, the two identical objects placed in the center of the arena. The two objects were positioned equidistant from the center of the arena, and equidistant from the walls of the arena. Each mouse was allowed to explore the objects for 10 minutes. Animals were then removed from the testing arena and placed in the home cage for 5 minutes. During this time, the arena was cleaned with 70% ethanol, and one of the objects was replaced with a novel object. The mouse was then placed back into the chamber and allowed to explore objects for 10 minutes (testing phase). The amount of time the mouse spent physically investigating (touching) each of the objects was assessed during both the learning and testing phases. If an animal did not physically touch both objects during the learning phase, it was excluded from NORT analysis.

Animals were also subjected to a 'control' NORT on the third day, where the test objects differed in color and shape, but not texture (5 minute shape NORT). In the learning phase of this assay, the animal was presented with two objects of identical texture, color and shape and was allowed to freely explore the objects for ten minutes. After a 5 minute retention period in the home cage, the animal was placed back into the test chamber, with one of the objects exchanged for a novel object of a different color and shape. Since mice prefer novel stimuli, an animal that can discriminate between the textures on the objects spends more time investigating the novel object (Heyser and Chemero, 2012), whereas an animal that does not discriminate novelty is expected to investigate the objects equally.

On the fourth day, a texture-specific NORT (textured NORT) was performed: in the learning phase, the animal was presented with two cubes of identical texture and was allowed to freely explore the objects for ten minutes. After a 5 minute retention period in the home cage, the animal was placed back into the test chamber, with one of the objects exchanged for a novel textured object. Since mice prefer novel stimuli, an animal that can discriminate between the textures on the objects spends more time investigating the novel textured object (Heyser and Chemero, 2012), whereas an animal that cannot discriminate between the textures is expected to investigate the objects equally (Orefice et al., 2016).

Lastly, a second control NORT was implemented on the fifth day, but with a one hour retention period between exploration and testing phases to assess memory performance (1 hour retention NORT).

For textured NORT, one object was replaced with a second, novel object that differed only in texture. Textured objects (either smooth or rough) were 4 cm on each side and constructed of plexiglass. All sides of the textured objects had ridges etched into the plexiglass using a laser cutter. For rough objects these ridges faced outwardly, while for smooth objects, the ridges pointed inward so that the objects appear visually identical but differed only in their textures. For the 5 minute shape and 1 hour retention versions of NORT, small wooden blocks were utilized. Blocks differed in shape, size and color, but overall volumes were similar (~6 cm x 3 cm x 3 cm). For each test, the placement and object used as the novel object was pseudorandomized and counterbalanced between groups.

To avoid confounding whisker movements and sensations, whiskers were plucked three days prior to the start of habituation (see above section on whisker plucking). Under these conditions, animals showed no adverse behavioral effects and used glabrous skin on their paws during more than 90% of object exploration (Orefice et al., 2016).

The position of the mouse was tracked using custom MATLAB scripts. Whisking, nose pokes and investigation using the paws were all included in the time spent investigating objects, though over 90% of the time investigating objects was performed using the glabrous skin on paws (Orefice et al., 2016). All three versions of NORT were analyzed using the same object exploration criteria. All groups exhibited a preference for the novel object during the 5 minute color/shape NORT, and the animal numbers per group are the same as the animal numbers for the textured NORT.

Startle Reflex/ Prepulse Inhibition (PPI)

PPI testing was performed as previously described (Orefice et al., 2016). The response of mice to tactile and acoustic startle stimuli, and the ability of a tactile or acoustic stimulus to inhibit startle to a loud acoustic stimulus was measured using a San Diego Instruments startle reflex system (SR-LAB Startle Response System). While the startle response to tactile stimuli has been previously investigated (Logue et al., 1997; Taylor et al., 1991; Torkamanzehi et al., 2008), we sought to determine whether mice exhibit specific tactile sensorimotor gating deficits by employing a PPI assay in which the pre-stimulus is an air puff (0.9 PSI, 50ms), followed by a startle tone stimulus (125 dB, 20ms). Air puffs were administered to the back of the mouse to assess hairy skin sensitivity. The 0.9 PSI prepulse stimulus strength was chosen because control animals showed a minimal, but statistically significant response to the stimulus alone, compared to baseline movement in the chamber without any stimulus (average response in controls, 5.89% \pm 2.15). In addition, this prepulse intensity was chosen because it produced significant inhibition of startle reflex at multiple interstimulus intervals (ISIs). A tone pre-stimulus (ranging from 68 dB to 80 dB, for 20ms), followed by startle tone stimulus (120 dB, 20ms) version of the PPI assay (acoustic PPI) was done as a control.

For testing sessions, animals were placed into a ventilated, cylindrical holder on a platform within a soundproof chamber. Protocols consisted of an acclimation phase, block I, block II, block III and block IV trials. Each time an animal was tested, it first underwent an acclimation phase to acclimate to the animal holder, startle box and background noise. Each mouse was placed in the chamber for a 5 minute acclimation period, during which constant background noise of broadband white noise was presented. Background noise for the acoustic PPI testing sessions was 65 dB. Background noise for the tactile PPI testing sessions was increased to 75 dB, to ensure that that the animal could not hear the air puff prepulse.

Block I consisted of 5 startle stimuli alone (120 or 125 dB broadband white noise for acoustic and tactile PPI testing sessions, respectively), to measure the initial startle reflex. Block II consisted of 5 prepulse stimuli alone (either 80 dB broadband white noise or a 0.9 PSI air puff for acoustic and tactile PPI testing sessions, respectively), to measure response to the prepulse stimulus alone. Block III incorporated prepulse/pulse, pulse alone and no stimulation trials that were pseudorandomized. Block IV consisted of 5 startle stimuli alone, to measure habituation to the startle stimuli over the testing session. Inter-trial intervals were varied from 10 s to 50 s (average 30 s). Acoustic PPI and tactile PPI sessions were run on separate days.

For acoustic PPI, the prepulse was 20ms in duration and presented 100 ms before the startle pulse (100 ms ISI). For tactile PPI, the prepulse intensity remained constant (0.9 PSI, 20 ms), and the ISI was varied from 50 ms to 1 s in duration. Whole body flinch, or startle reflex, was quantified using an accelerometer sensor measuring the amplitude of movement of the animal within the cylindrical holder.

Elevated Plus Maze

The elevated plus maze was used to measure anxiety-like behavior (Moy et al., 2007). The elevated plus maze was a custom-built acrylic chamber. The arms were each 30 cm long x 5 cm wide, with two of the opposing arms having walls that were 15 cm high. The maze stood 40 cm above the ground. Testing occurred in a dimly lit room with a camera overhead for tracking animal movement. The animal was placed into the center of the maze and allowed to freely explore for 10 minutes. The time spent in either the center, closed arms or open arms of the maze during each test session was quantified using a custom MATLAB script. The percentage of time each mouse spent in the open arms, compared to the closed arms and center of the chamber was calculated and expressed as a percent time spent in the open arms of the maze.

3-Chamber Social Interaction Test

The 3-chamber social interaction assay is a commonly used method of assessing both sociability and social recognition/preference in rodents, and is often used to measure ASD-like behavioral deficits in mice (Moy et al., 2007). Testing occurred in three sessions within a three-chambered box with openings between the chambers (each compartment is 20 cm wide x 40 cm long x 22 cm high). The outside walls of the chamber were opaque, while the inner dividers were clear plexiglass. After a 5 minute habituation period in the empty chamber, the test mouse was moved into the empty center chamber with partitions in place. A wire mesh cup containing a novel mouse was placed on one side of the chamber and an empty mesh cup was placed on the other side during the “sociability” session. The partitions were then lifted, and the test mouse was free to explore all three sections of the chamber. After 10 minutes, the test mouse was moved into the empty center chamber with partitions in place. A second novel mouse was placed under the empty cup, partitions were removed, and the test mouse was allowed to freely explore the chamber during the social novelty recognition preference session (“Social Novelty Preference”) for 10 minutes. The time spent in each of the three chambers during each test session was quantified using custom MATLAB scripts. Control mice show a significant preference for the chamber containing the novel mouse in each portion of the test, indicating a preference for novel mice compared to novel objects as well as a preference (and recognition of) a novel mouse versus a familiar mouse.

Marble Burying

The marble burying test is used to measure both restricted repetitive behaviors and neophobia in mice (Jaramillo et al., 2017). Each mouse is placed into the center of a separate, clean cage with 2 inches of normal bedding and 12 marbles on top of the bedding arranged in three even rows. Mice are left alone in these individual cages for 20 minutes, during this time mice typically explore the new environment and dig in the bedding ultimately burying marbles in the process. After 20 minutes, the experimenter re-entered

the room and removed each mouse from its testing cage. Images of each cage were acquired, and the number of marbles buried (denoted by greater than 2/3 of the marble not being visible) was quantified per mouse.

Phenotypic Scoring

Mice were observed for indicators of general well-being such as coat condition, eyes, and body stance as previously described (Guy et al., 2007). A score of '0' indicated a clean shiny coat, clear eyes and normal stance. A score of '1' indicated the appearance of dull eyes, dull/ungroomed coat and somewhat hunched stance. A score of '2' denoted narrowed or crusted eyes, piloerection and a hunched posture. A score of '3' indicated severe eye narrowing, profound piloerection or loss of fur, and severely hunched posture.

Immunohistochemistry

For histology, mutant and control littermates were sacrificed in pairs or groups. Mice (P42-P84) were anesthetized with isoflurane and transcardially perfused with 20 mL of PBS, following by 25 mL of 4% paraformaldehyde (PFA) in PBS at room temperature (RT). Brains, spinal cords and dorsal root ganglia (DRG) were dissected from perfused mice and post-fixed overnight in 4% PFA at 4°C. Tissue was then washed 3 times in 1X PBS for 1-2 hours each, then brain, spinal cord and attached DRGs were finely dissected out of the vertebral column. Tissue was cryoprotected in 30% sucrose in 1X PBS overnight at 4°C, then embedded in NEG50 and frozen at -20°C. Tissue was stored at -70°C until it was sectioned at 30 µm using a cryostat.

Sections on slides were dried at RT for 1 hour, then washed 3x10 minutes in 1X PBS. Tissue was then blocked for 1-2 hours in 1X PBS containing 0.3% Triton X-100 and 5% normal donkey or goat serum (Jackson Immuno, 005-000-121). Sections were incubated with primary antibodies diluted in blocking solution (5% normal donkey serum, no detergent) at 4°C for 1-2 days. After which, sections were washed with 1X PBS containing 0.02% Tween 20 (PBST) and then incubated with secondary antibodies diluted 1:500 in blocking solution, at room temperature for 2 hours. IB4 was diluted at 1:500 and applied with secondary antibodies when needed. Following secondary antibody incubation, tissue was washed 3x10 minutes with PBST. One wash of PBST contained a 1:10,000 dilution of Hoechst solution. Sections were washed one more time with PBST, then mounted with fluoromount-G (Southern Biotech). Primary antibodies used: mouse anti-Shank3 (ab193307, Abcam, 1:1000); guinea pig anti-vGLUT1 (AB5905, Millipore, 1:1000); rabbit anti-HCN1 (APC-056, Alomone Labs, 1:500); rabbit anti-KCNQ2 (ab22897, Abcam, 1:500); goat anti-GABRB3 (PA5-19060, Life Technologies, 1:250); rabbit anti-PARVALBUMIN (PV-27, Swant, 1:1000); goat anti-PARVALBUMIN (PVG-213, Swant, 1:1000); rabbit anti-MECP2 (provided by Michael Greenberg, 1:1000); mouse anti-GAD67 (MAB5406, Fisher Scientific, 1:1000); rabbit anti-dsRed (632496, Clontech, 1:1000); and Alexa 647-conjugated IB4 (I6520-100, Life Technologies, 1:1000). Secondary antibodies include: Alexa 488, 546 or 647 conjugated donkey or goat anti-mouse, rabbit, goat or guinea pig. All secondary antibodies were purchased from Life Technologies or Jackson Immuno Research and used at a 1:500 dilution.

Puncta Analysis

Z stack images of spinal cord slices were taken on a Zeiss LSM 700 confocal microscope using a 63X oil-immersion lens (Zeiss; Plan-Apochromat 63X/NA 1.40). Images were taken in lamina III/IV of the dorsal horn, which was identified by immunostaining with IB4 to delineate lamina II. The percent of SHANK3, GABRB3, HCN1 or KCNQ2 puncta that were colocalized with vGLUT1 puncta were analyzed using NIH ImageJ software with a custom ImageJ plugin (Schneider et al., 2012). Colocalization was analyzed per 0.5-micron section thickness of tissue, with a total of ten continuous sections. At least three images from three to seven animals per genotype were analyzed for each experiment. Experimenters were blind to genotype and treatment conditions during both experiment and analysis portions of the experiments.

Parvalbumin⁺ Neuron Analyses

Z stack images of coronal brain sections were taken on a Zeiss LSM 700 confocal microscope using a 63X oil-immersion lens (Zeiss; Plan-Apochromat 10X/NA 0.45 and 20X/NA 0.8). Images were taken in trunk primary somatosensory cortex (S1), primary visual cortex (V1), and basolateral amygdala (BLA). Cell bodies of parvalbumin-positive (PV⁺) neurons in each of these regions were traced using ImageJ. The number, size and fluorescence intensity of PV⁺ neurons in 1 mm² region were analyzed per image, and 2-3 images were analyzed per region of interest. Four to seven animals were analyzed per genotype per experiment, by an experimenter blind to both genotype and treatment conditions.

Cultured Dorsal Root Ganglia Electrophysiology

6-8 week old mice were euthanized via CO₂ narcosis, and dorsal root ganglia (DRGs) were rapidly dissected out on ice. DRGs were digested in collagenase (2 mg/mL)/dispase (5 mg/mL) solution at 37°C with constant rotation for 45 minutes. Following enzymatic digesting, tissue was centrifuged, washed three times with 1X HBSS and then mechanically triturated using a glass Pasteur pipette. Dissociated cells were plated onto coverslips coated with poly-D-lysine (50 µg/mL) and laminin (1 µg/mL). Cells were allowed to attach to coverslips for 30 minutes, and then high glucose DMEM supplemented with penicillin, streptomycin and fetal bovine serum was added to each well. Patch-clamp recording of DRG neurons was performed 1-2 days post plating.

Coverslips with attached, cultured dorsal root ganglion neurons were stored in a submerged recording chamber at room temperature and continuously perfused with extracellular solution containing 132 mM NaCl, 3 mM KCl, 2.5 mM CaCl₂, 10 mM HEPES, and 10 mM glucose, saturated with 95% O₂, 5% CO₂ at a rate of ~2 ml/min (pH 7.4). Cells were visualized using infrared differential

interference contrast microscopy. Whole cell voltage-clamp and current-clamp recordings from neurons were obtained under visual guidance using a 40x objective. Patch electrodes (3.0–4.0 M Ω) were filled with a KCl-based internal solution containing 134 mM KCl, 1 mM MgCl₂, 1 mM CaCl₂, 10 mM HEPES, 10 mM EGTA, 4 mM ATP-Mg₂₊ and 0.4 mM GTP-Na⁺ (pH 7.4, 300 mOsm). Neurons were voltage clamped and subjected to a hyperpolarization voltage-step protocol to assess I_h and I_m. Neurons were held in current clamp and excitability was assessed using increasing steps of current (100 pA) to determine the threshold at which each neuron would fire an action potential. In a subset of experiments, 10 μ M isoguvacine was bath applied and the same experiments were repeated. At the end of an experiment, 100 μ M ZD-7288 was bath applied, followed by bath application of 5mM CsCl₂, to block HCN channels. Images of each recorded neuron were acquired and the size of each cell body was recorded. Data were acquired using a Multiclamp amplifier, a Digidata 1440A acquisition system, and pClamp10 software (Molecular Devices). Sampling rate was 10 kHz. No correction for junction potential was applied. Cells were discarded if membrane potential (V_m) at break-in was > -60 mV, R_{in} was < 75 M Ω , residual uncompensated R_s was > 20 M Ω . Input resistance and access resistance were monitored continuously throughout each experiment and cells were excluded from analysis if these values changed by more than 10% during the experiment.

For each culture and recording session, a control and mutant littermate were prepared at the same time, with the experimenter blind to genotype. Electrophysiology recordings alternated between coverslips of the two different conditions for the duration of the experimental session. The experimenter was blind to genotype and treatment conditions during both experiment and analysis portions of the experiments.

Brain Slice Electrophysiological Recordings

8–10 week old mice were deeply anesthetized and rapidly transcardially perfused with choline chloride solution (92 mM choline chloride, 2.5 mM KCl, 1.2 mM NaH₂PO₄, 30 mM NaHCO₃, 20 mM HEPES, 25 mM glucose, 5 mM sodium ascorbate, 2 mM thiourea, 3 mM sodium pyruvate, 10 mM MgSO₄·7H₂O, 0.5 mM CaCl₂·2H₂O). Coronal brain sections (300 μ m) were cut anterior to posterior, including sections containing primary somatosensory cortex and primary visual cortex. Slices recovered for 25 minutes at 35°C in HEPES recovery solution (76 mM Tris-HCl, 19.5 mM Tris base, 2.5 mM KCl, 1.2 mM NaH₂PO₄, 30 mM NaHCO₃, 20 mM HEPES, 25 mM glucose, 5 mM sodium ascorbate, 2 mM thiourea, 3 mM sodium pyruvate, 10 mM MgSO₄·7H₂O, 0.5 mM CaCl₂·2H₂O). After 25 minutes at 35°C, slices were kept at room temperature in HEPES recovery solution until recording experiments began. Brain slices were stored in a submerged recording chamber at room temperature and continuously perfused with ACSF containing 2.5 mM CaCl₂·2H₂O, 1 mM NaH₂PO₄·H₂O, 119 mM NaCl, 2.5 mM KCl, 1.3 mM MgSO₄·7H₂O, 26 mM NaHCO₃, 25 mM dextrose, and 1.3 mM Na L-ascorbate (pH 7.4, 305–310 mOsm), saturated with 95% O₂, 5% CO₂ at a rate of ~2 ml/min. Cells were visualized using infrared differential interference contrast microscopy. Whole cell voltage-clamp and current-clamp recordings from layer 2/3 pyramidal neurons in trunk S1 or V1 were obtained under visual guidance using a 40x objective. Patch electrodes (3.5–4.5 M Ω) were filled with a CsCl-based internal solution containing 117 mM CsCl, 87 mM CsOH, 2.8 mM NaCl, 5 mM MgCl₂, 2 mM ATP-Na⁺, 0.3 mM GTP-Na⁺, 0.6 mM EGTA, and 20 mM HEPES (pH 7.25–7.28, 275–285 mOsm), and neurons were voltage clamped at -70 mV and then at 0 mV. Spontaneous excitatory postsynaptic currents (sEPSCs, -70 mV hold) and inhibitory postsynaptic currents (sIPSCs, 0mV hold) were each measured during a 5–10 minute period. At the end of an experiment, 2,3-dihydroxy-6-nitro-7-sulfamoyl-benzof[*q*]quinoxaline (NBQX; 10 μ M) was bath-applied to block AMPA receptors, followed by bath-applied 3 μ M gabazine to block GABA_A receptors. For a separate set of neurons, patch electrodes (3.5–4.5 M Ω) were filled with a K-gluconate-based internal solution containing 150 mM K-gluconate, 3 mM KCl, 3 mM ATP-Mg₂₊, 0.5 mM GTP-Na⁺, 0.5 mM EGTA, 5 mM phosphocreatine-tris₂, 5 mM phosphocreatine-Na₂ and 10 mM HEPES (pH 7.2, 290–300 mOsm), and held in current clamp to confirm excitatory pyramidal neuron firing properties. Data were acquired using a Multiclamp amplifier, a Digidata 1440A acquisition system, and pClamp10 software (Molecular Devices). Sampling rate was 10 kHz, and data were low-pass filtered at 3 kHz. No correction for junction potential was applied. Cells were discarded if membrane potential (V_m) at break-in was > -50 mV, or R_{in} was < 75 M Ω , residual uncompensated R_s was > 20 M Ω . Input resistance and access resistance were monitored continuously throughout each experiment and cells were excluded from analysis if these values changed by more than 10% during the experiment. Four animals were included per genotype per experiment, with 2–4 neurons analyzed per brain region, per mouse (S1 or V1). E/I ratios were calculated as the frequency of spontaneous ePSCs divided by the frequency of spontaneous iPSCs per cell. Averages per brain region, per animal, were calculated as the average of the E/I ratios per cell analyzed. The experimenter was blind to genotype and treatment conditions during both experiment and analysis portions of the experiments.

In Vivo DRG Multi-Unit Electrode Recordings

In vivo recordings were performed on mutant mice and their control littermates at 8–12 weeks of age, using a preparation modified from prior studies (Ma and LaMotte, 2007). This preparation allows measurement of action potentials evoked by cutaneous stimulation for up to four hours *in vivo*. Mice were administered dexamethasone 1 hour prior to the start of surgery. Each animal was anesthetized using a combination of urethane (1mg/g) and isoflurane (2%), and then surgery was performed to expose the left, L4 DRG. Temperature was monitored and maintained at 35.5–37.5°C with a temperature controller (TC-344B, Warner Instruments) and thermoelectric heater (C3200-6145, Honeywell) embedded in castable cement (Aremco). At the beginning of the DRG exposure, an incision was made over the spine (T10 - L6) and the overlying tissue retracted to expose the vertebral column. The spine was secured with custom spinal clamps (Mike's Machine, Attleboro MA) and bone dorsal to the target DRG removed with rongeurs.

Following surgery, and during the entire recording preparation, animals were maintained on 1% isoflurane. A NeuroNexus tetrode was inserted into the DRG under visual guidance using a 10x objective. Light touch responsive units were identified as units that fired action potentials in response light brush and air puff stimuli. Units were characterized as glabrous or hairy skin innervating A β RA1-LTMRs, A β SA1-LTMRs or A β field-LTMRs, or proprioceptors, based on firing rate, and optimal stimulus type (Figures 6D and 6E). In this preparation, multiple single units (2-12 per experiment; example of typical single unit in Figures 6B and 6C) with receptive fields on the thigh or hind paw could be identified following skin stimulation with innocuous mechanical stimuli.

Once LTMR units were identified, animals were administered a subcutaneous injection of either saline vehicle or isoguvacine (2 mg/kg), and sensitivity and firing frequency in response to tactile stimuli were monitored over a 90-minute period for each identified unit. Voltage at each tetrode channel was bandpass filtered (1 to 7500 Hz), amplified, and digitized at 20 kHz with an Intan RHD2132 chip attached to an Intan RHD2000 Evaluation System. Brushing was monitored with a strain gauge (MBL (BL341AH) 25 gram Model MBL load cell, Sensotec-Honeywell) connected to an amplifier (DMD465WB, Omega). Data from the strain gauge were digitized at 20 kHz by a 16-bit A/D converter (USB-6259, National Instruments). Spikes were detected from the voltage data and sorted using JRClust (Figures 6B–6E) (Jun et al., 2017). Firing properties were analyzed using custom code (Python 3.6). The experimenter was blind to genotype and treatment conditions during both experiment and analysis portions of the experiment.

Liquid Chromatography Mass Spectrometry

All samples were harvested from 8-12 week old male mice. For cerebrospinal fluid (CSF) and plasma sample preparation: 5 μ L of sample (if less was present, water was added to reach 5 μ L and the concentrations were calculated appropriately) was mixed with 20 μ L of a 150 nM solution of nipecotic acid in methanol (Sigma Aldrich). The samples were incubated at -20°C for 2 hours to precipitate the proteins. After centrifugation at 10000 g for 35 minutes, the supernatants were transferred to microinserts, dried under nitrogen flow and resuspended in 15 μ L of water. A set of calibration standards (1/5 dilution series, starting at 50 μ M) is prepared the same way as the fluid samples.

Tissue sample preparation: weighed pieces of frozen tissue were transferred to bead beater tube containing 1mL of 50 nM nipecotic acid (internal standard) in cold methanol and garnet pieces (SPEX SamplePrep tubes). Samples were homogenized for 10 minutes at 50 Hz in a TissueLyser LT (Biorad). Homogenized samples were transferred to 8 mL glass vial. The Bead Beater vials were washed with 1 mL methanol, and the methanol were combined with the samples. 4 mL of chloroform was added and the samples were incubated 1 hour at room temperature. Phase separation was then initiated by adding 2 mL water to each sample. After vortexing, the samples were centrifuged at 2500 g for 50 minutes at 4°C . The aqueous phases were then transferred to new vial, dried under nitrogen flow, and resuspended in 100 μ L of water. A set of calibration standards was also prepared using the method.

Isoguvacine concentration was quantified on a Thermo Q-Exactive plus mass spectrometer coupled with an Ultimate 3000 LC (both Thermo Fisher). 8 μ L of fluid samples or 5 μ L of tissue samples were injected on a PFPP column (Phenomenex, 150x2mm). Mobile phase A was water with 1% acetic acid and mobile phase B was Acetonitrile with 1% acetic acid. The elution gradient was as follow: 3min with 0% B, then to 100%B in 2 min, followed by 3 minutes at 100%B. the column was then re-equilibrated with 0% B for 4 min. The flow rate was 0.2 mL/min. The MS was run in positive ion mode at 70000 resolution in full MS and in PRM mode. Quantification was achieved by integrating the signal of the transition 128.07061 to 82.06570 for isoguvacine, 130.08626 to 112.0760 for Nipecotic acid (internal standard), all at a normalized collision energy of 80 and a 1 m/z isolation window. A ten-point calibration curve was used ranging from 25 pM to 50 μ M. The detection limits for isoguvacine were 640 pM in the fluid samples and 16 nM in the tissue samples, and 128 pM for diazepam. Blank samples were run in between each sample to insure no carry over interferences.

GABA_A Receptor Agonist Assay

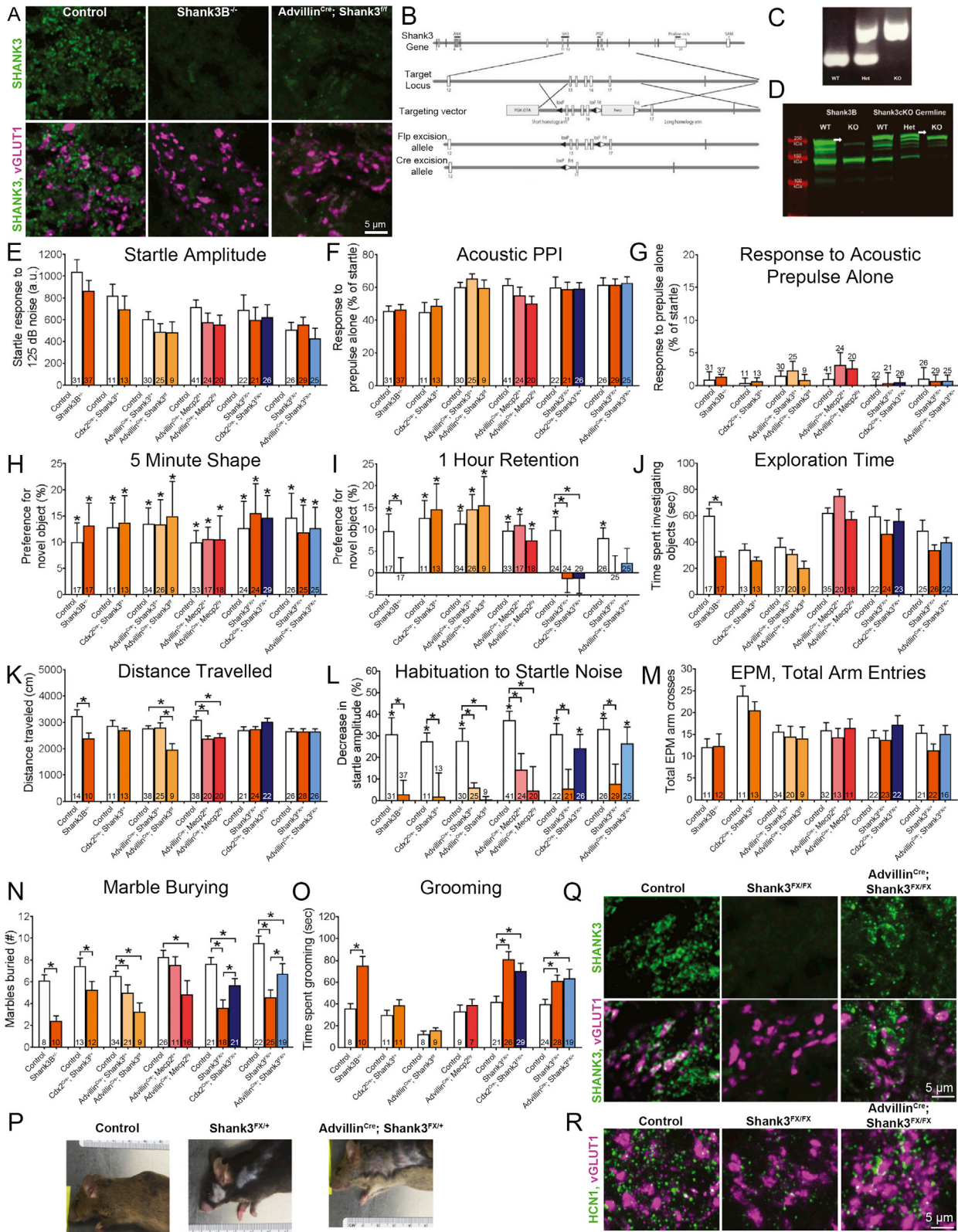
Isoguvacine agonism for the GABA_A receptor was tested using the IonFlux HTTM Automated Patch Clamp System (Eurofins). HEK293T cells transfected with human GABA_A α 1 β 3 γ 2 or α 21 β 3 γ 2 subunits were subjected to patch clamp electrophysiology in the presence of varying concentrations (6-point concentration response) of isoguvacine for two seconds during each application. The maximal Cl⁻ ion current elicited during each drug application was recorded. Six-point concentration response of GABA was tested to serve as an agonist control. Eight replicates were performed for each drug at each concentration. DMSO application for 2 s served as a time matched vehicle control. Peak isoguvacine-evoked ion currents were normalized to the peak Cl⁻ ion current evoked by EC₁₀₀ GABA for 2 s.

QUANTIFICATION AND STATISTICAL ANALYSIS

For all figures and tables, data are expressed as the mean values \pm standard error of the mean (SEM). Bartlett's test for equal variance was applied to all datasets, and the variances among groups were not shown to be statistically different from each other. The number of animals per group used in each experiment is denoted within the bar for that group in each panel.

For NORT, three-chamber social interaction tests, and habituation to startle noise experiments, each group was analyzed compared to a chance level using a one sample t test (chance level: 0% preference or 0% habituation), with statistical significance being denoted as a symbol directly above the bar being analyzed. Comparisons between groups in all experiments were performed using Student's t test (in the case of two groups in one condition), one-way ANOVA (in the case of three or more groups in one condition), or two-way

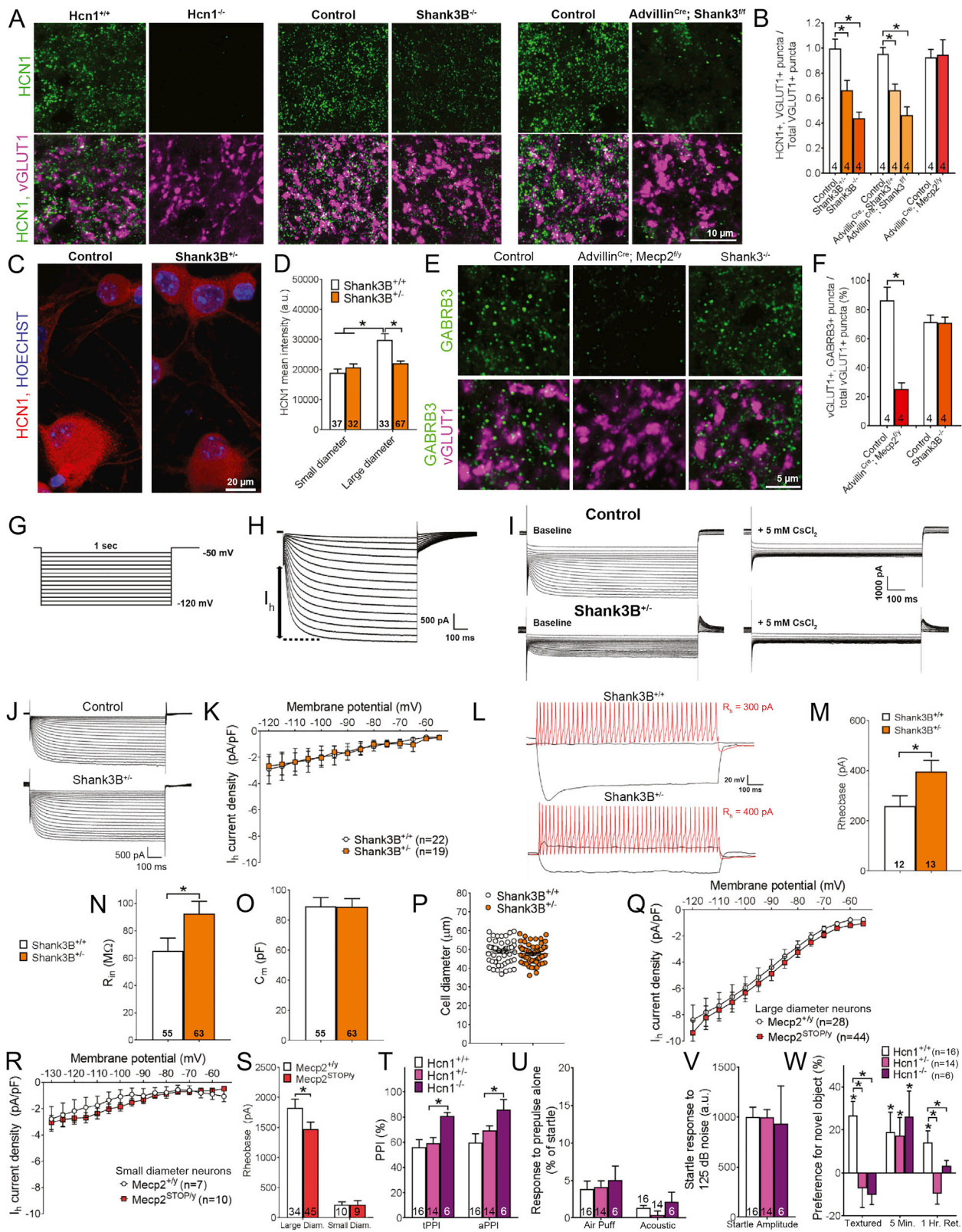
ANOVA (in the case of at least two groups with multiple conditions or time points). Comparisons between groups with significant differences are indicated using a bracket above the appropriate groups. Asterisks indicate p values for Student's t tests, unless otherwise noted in the figure legends. Unless otherwise noted, *, $p < 0.05$ and #, $p < 0.10$. Main effects from one-way and two-way ANOVAs are expressed as an F-statistic and P value within brackets of the figure or denoted in Supplemental Tables. For one-way and two-way ANOVAs, post hoc comparisons were performed using the post hoc test indicated in the figure legend. The p values of post hoc comparisons are represented with asterisks in figures. All statistics were performed using GraphPad Prism.



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Figure S1. *Shank3* Functions Cell-Autonomously in Peripheral Somatosensory Neurons for Normal Innocuous Touch Behaviors, Related to Figure 1

- (A) IHC images of spinal cord (SC) dorsal horn lamina III/IV from control, *Shank3B*^{-/-} or *Advillin*^{Cre}; *Shank3*^{fl/fl} mice, showing SHANK3 expression at vGLUT1+ presynaptic terminals for A β and A δ LTMRs.
- (B) Schematic describing strategy to create *Shank3B* conditional knockout (*Shank3*^f) mouse. Mice containing targeted allele were crossed to Flp mice to remove the Neo cassette and β -*Actin*^{Cre} mice to excise Exons 13-16.
- (C) PCR genotyping confirms successful deletion of Exons 13-16 from genome of *Shank3*^f.
- (D) Western blot of whole brain lysates from *Shank3B*^{-/-} and β -*Actin*^{Cre}; *Shank3*^f mice shows deletion of protein products from full-length isoform (white arrows).
- (E) Magnitude of startle response to a 125 dB noise in mutant mice and control littermates. Student's unpaired t test or one-way ANOVA, not significant.
- (F) Percent inhibition of the startle response to a 125 dB noise (pulse), when the startle noise is preceded by tone prepulse (80 dB, 'acoustic PPI') in mutant mice and control littermates.
- (G) Response to a non-startling acoustic noise (80 dB, 20 ms), mutant mice and their control littermates. Responses are expressed as percent of startle response to a 125 dB startle noise. Student's unpaired t test or one-way ANOVA, not significant.
- (H) Discrimination index for 5 minute shape NORT. Student's unpaired t test or one-way ANOVA, not significant.
- (I) Discrimination index for 1 hour retention NORT. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (J) Average amount of time (seconds) spent physically interacting with both the familiar and novel object in the NOR tests in mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (K) Average total distance traveled in the open field chamber for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (L) Percent decrease in startle response to a 125 dB noise during a 30-minute tactile PPI session, when comparing the first five startle responses to the last five responses to a 125 dB noise for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (M) Average number of EPM arm entries during the 10-minute assay, for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA, not significant.
- (N) Average number of marbles buried (out of 12) during a twenty-minute assay for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (O) Average amount of time spent grooming during a 10-minute open field assessment for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.
- (P) Representative images of control, *Shank3*^{FX/+} and *Advillin*^{Cre}; *Shank3*^{FX/+} mice, showing excessive grooming and loss of fur in the *Shank3*^{FX/+} and *Advillin*^{Cre}; *Shank3*^{FX/+} mice.
- (Q) IHC images of SC dorsal horn lamina III/IV from control, *Shank3*^{FX/FX} or *Advillin*^{Cre}; *Shank3*^{FX/FX} mice, showing SHANK3 expression at vGLUT1+ presynaptic terminals.
- (R) IHC images of SC dorsal horn lamina III/IV from control, *Shank3*^{FX/FX} or *Advillin*^{Cre}; *Shank3*^{FX/FX} mice, showing HCN1 expression at vGLUT1+ presynaptic terminals. Error bars indicate standard error of the mean.



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Figure S2. Loss of *Shank3* Leads to Increased Excitability of Large Diameter Peripheral Somatosensory Neurons, Related to Figure 1

- (A) IHC images of SC dorsal horn lamina III/IV from *HCN1*^{-/-}, *Shank3B*^{-/-}, *Advillin*^{Cre}; *Shank3*^{fl/fl} mice and their control littermates, showing HCN1 expression at vGLUT1+ presynaptic terminals.
- (B) Quantification for the number of HCN1 puncta that co-localize with vGLUT1+ puncta in lamina III/IV of the spinal cord dorsal horn of mutants and control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, [for germline *Shank3* mutants: F (2,6) = 24.76, p = 0.0013; for conditional *Shank3* mutants: F (2,6) = 14.25, p = 0.0053], *p < 0.05.
- (C) Immunocytochemistry images of cultured DRG neurons from *Shank3*^{+/-} mice and their control littermates, showing HCN1 expression.
- (D) HCN1 expression in small diameter (< 30 μm diameter) and large diameter (> 40 μm diameter) cultured DRG neuron cell bodies, for control and *Shank3*^{+/-} mutant mice. One-way ANOVA with post hoc Tukey's test [F (2,165) = 5.868, p = 0.0165], *p < 0.05.
- (E) IHC images of spinal cord (SC) dorsal horn lamina III/IV from *Advillin*^{Cre}; *Mecp2*^{fl/y} and *Shank3*^{-/-} mice and their control littermates, showing GABRB3 expression at vGLUT1+ presynaptic terminals.
- (F) Quantification for the number of GABRB3 puncta that co-localize with vGLUT1+ puncta in lamina III/IV of the spinal cord dorsal horn of mutants and control littermates. Student's unpaired t test, *p < 0.05.
- (G) Voltage step protocol used to activate HCN channels and elicit I_h during whole-cell voltage clamp recordings.
- (H) Whole-cell voltage clamp trace showing I_h during a hyperpolarizing voltage step protocol in a large diameter neuron cultured from mouse DRGs, which is blocked following application of 5 mM CsCl₂.
- (I) Representative traces from large diameter DRG neurons cultured from a control and *Shank3B*^{+/-} mice, showing I_h during a hyperpolarizing voltage step protocol, with and without application of CsCl₂ to block HCN channels.
- (J) Representative traces from small diameter DRG neurons cultured from a control and *Shank3B*^{+/-} mice, showing I_h during a hyperpolarizing voltage step protocol.
- (K) Quantification of total I_h density at each hyperpolarizing voltage step for small diameter neurons cultured from DRGs of control and *Shank3B*^{+/-} mutant mice. *Repeated-measures two-way ANOVA* [F (1,532) = 0.001563; p = 0.9685], not significant.
- (L) Representative traces from small diameter neurons cultured from DRGs of control and *Shank3B*^{+/-} mice during whole cell current clamp recordings, in which the minimal amount of current required to elicit an action potential in each neuron (rheobase, R_h), was determined.
- (M) Average R_h in small diameter neurons cultured from DRGs of control and *Shank3B*^{+/-} mice during whole-cell current clamp recordings. Student's unpaired t test, p < 0.05.
- (N) Average input resistance (R_{in}) of large diameter neurons cultured from DRGs of control and *Shank3B*^{+/-} mice during whole-cell patch clamp recordings. Student's unpaired t test, p < 0.05.
- (O) Average capacitance (C_m) of large diameter neurons cultured from DRGs of control and *Shank3B*^{+/-} mice during whole-cell patch clamp recordings. Student's unpaired t test, not significant.
- (P) Average cell body size of large diameter neurons cultured from DRGs of control and *Shank3B*^{+/-} mice during whole-cell patch clamp recordings. Student's unpaired t test, not significant.
- (Q) Quantification of total I_h density at each hyperpolarizing voltage step for large diameter neurons cultured from DRGs of control and *Mecp2*^{STOP/y} mutant mice. *Repeated-measures two-way ANOVA* [F (1,980) = 3.474, p = 0.0626], not significant.
- (R) Quantification of total I_h density at each hyperpolarizing voltage step for small diameter neurons cultured from DRGs of control and *Mecp2*^{STOP/y} mutant mice. *Repeated-measures two-way ANOVA* [F (1,240) = 3.147, p = 0.0773], not significant.
- (S) Average R_h in large and small diameter neurons cultured from DRGs of control and *Mecp2*^{STOP/y} mice during whole-cell current clamp recordings. Student's unpaired t test, p < 0.05.
- (T) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff ('tactile PPI'), or tone prepulse ('acoustic PPI') in mutant mice and control littermates. One-way ANOVA with post hoc Tukey's test [for tactile PPI: F (2,33) = 7.064, p = 0.0028; for acoustic PPI: F (2,28) = 3.999, p = 0.0297], *p < 0.05.
- (U) Response to a light air puff stimulus (0.9 PSI, 50 ms) or non-startling tone prepulse (80dB, 20 ms) alone. Responses are expressed as percent of startle response to a 125 dB noise. One-way ANOVA [F (2,33) = 0.2048, p = 0.8158], not significant.
- (V) Magnitude of startle response to a 125 dB noise in mutant mice and control littermates. One-way ANOVA [F (2,33) = 0.04956, p = 0.9517], not significant.
- (W) Discrimination index for textured NORT, 5 minute shape NORT and 1 hour retention NORT. One-way ANOVA with post hoc Tukey's test [for textured NORT: F (2,33) = 7.064, p = 0.0028; for 1 hour retention NORT: F (2,33) = 6.308, p = 0.0048], *p < 0.05. Error bars indicate standard error of the mean.

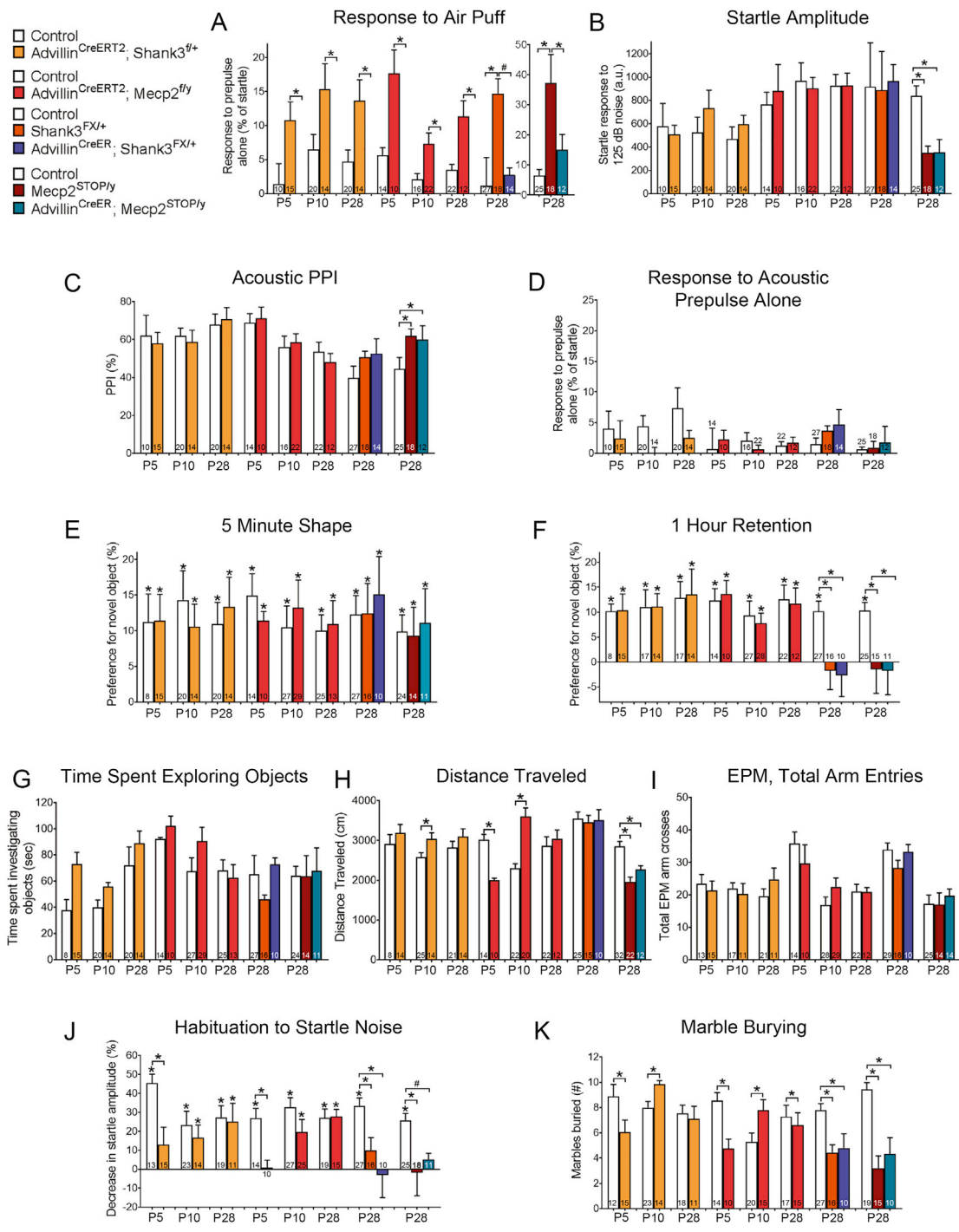


Figure S3. Normal Tactile Sensitivity Is Necessary during Early Postnatal Periods for Normal Behaviors in Adult Mice, Related to Figure 3

(A) Response to a light air puff stimulus alone mutant mice and their control littermates. Responses are expressed as percent of startle response to a 125 dB noise. Mice received five days of tamoxifen (TAM) treatment, beginning at either P5, P10 or P28. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05, #p < 0.10.

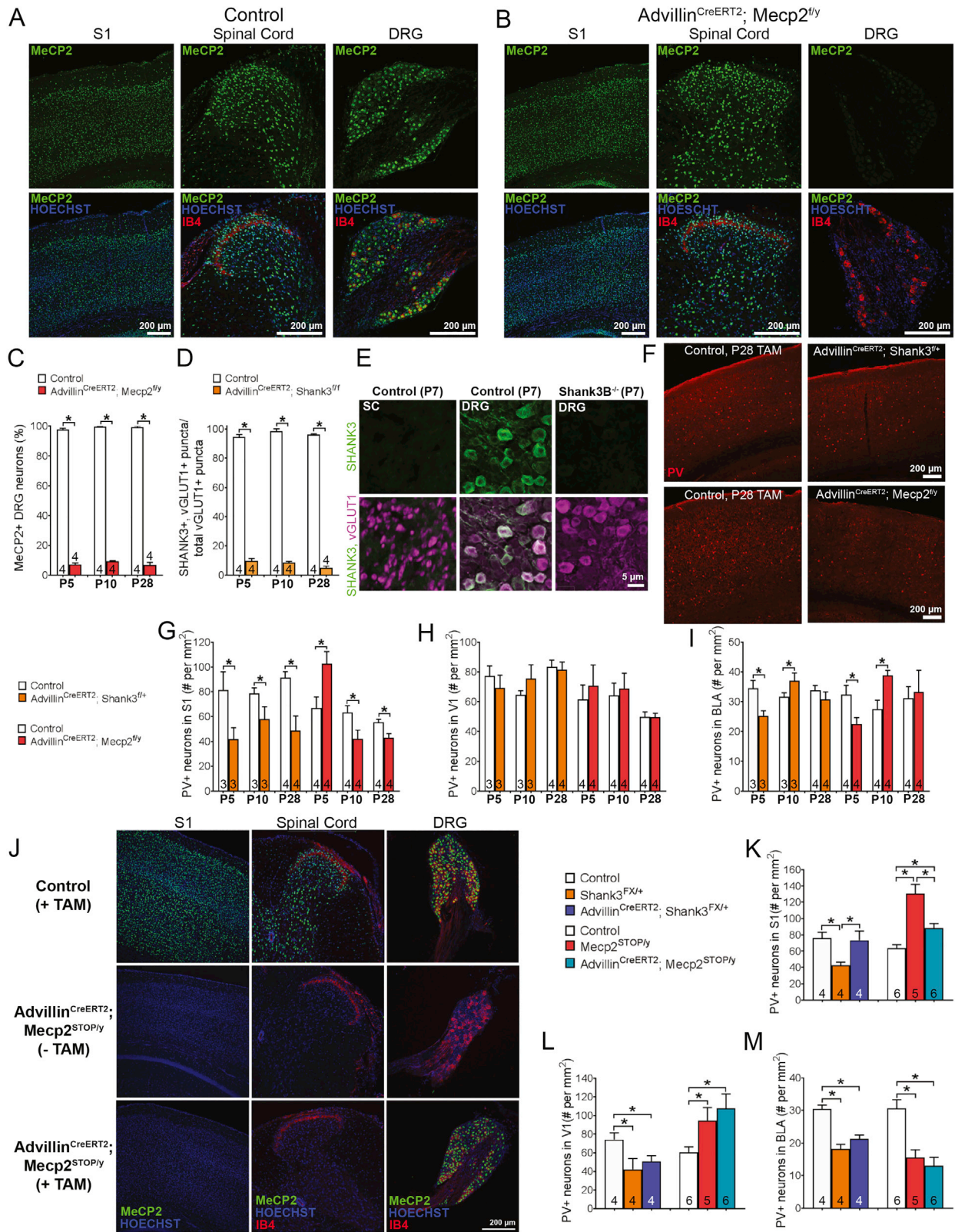
(B) Magnitude of startle response to a 125 dB noise in mutant mice and control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.

(C) Percent inhibition of the startle response to a 125 dB noise (pulse), when the startle noise is preceded by tone prepulse in mutant mice and control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, *p < 0.05.

(D) Response to a non-startling acoustic noise (80 dB, 20 ms) in mutant mice and their control littermates. Responses are expressed as percent of startle response to a 125 dB startle noise. Student's unpaired t test or one-way ANOVA, not significant.

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- (E) Discrimination index for 5 minute shape NORT. For comparisons between groups, Student's unpaired t test or one-way ANOVA, not significant.
- (F) Discrimination index for 1 hour retention NORT. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (G) Average amount of time (seconds) spent physically interacting with both the familiar and novel object in the NOR tests in mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA, not significant.
- (H) Average total distance traveled in the open field chamber for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (I) Average number of EPM arm entries during the 10-minute assay, for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA, not significant.
- (J) Percent decrease in startle response to a 125 dB noise during a 30-minute tactile PPI session, when comparing the first five startle responses to the last five responses to a 125 dB noise for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (K) Average number of marbles buried (out of 12) during a twenty-minute assay for mutant mice and their control littermates. Student's unpaired t test or one-way ANOVA with post hoc Tukey's test, * $p < 0.05$. Error bars indicate standard error of the mean.



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Figure S4. Loss of *Mecp2* or *Shank3* in Peripheral Somatosensory Neurons during Early Postnatal Periods Impacts Brain Development, Related to Figure 3

(A) IHC images of coronal primary somatosensory cortex (S1), spinal cord, or DRG sections from a control mouse showing MeCP2 expression (top panels) or MeCP2, Hoechst and IB4 expression (bottom panels).

(B) IHC images of coronal primary somatosensory cortex (S1), spinal cord, or DRG sections from an *Advillin^{CreERT2}; Mecp2^{fl/y}* mouse that received TAM treatment, showing MeCP2 expression (top panels) and MeCP2, Hoechst and IB4 expression (bottom panels). Note lack of MeCP2 expression in the DRG, while MeCP2 expression is normal in S1 and spinal cord sections.

(C) Average percentage of MeCP2+ DRG neurons in control mice or mice with sensory-neuron specific deletion of *Mecp2* (*Advillin^{CreERT2}; Mecp2^{fl/y}*), receiving five days of TAM beginning at P5, P10 or P28. Three to five DRGs were analyzed per mouse, and three to five mice per genotype and TAM date were included in the analysis. Student's unpaired t test, * $p < 0.001$.

(D) Average percentage of SHANK3+, vGLUT1+ puncta in spinal cord sections from control mice or mice with sensory-neuron specific deletion of *Shank3* (*Advillin^{CreERT2}; Shank3^{fl/fl}*), receiving five days of TAM beginning at P5, P10 or P28. Three to five spinal cord sections were analyzed per mouse, and three to five mice per genotype and TAM date were included in the analysis. Student's unpaired t test, * $p < 0.001$.

(E) IHC images of transverse spinal cord (SC) and DRG sections from a control mouse showing SHANK3 expression (top panels) or SHANK3 and vGLUT1 expression (bottom panels) in control mice. Note lack of SHANK3 expression in control SC at P7, and robust SHANK3 expression in DRG at P7.

(F) Representative IHC images of coronal S1 brain sections, showing PV immunoreactivity in control, *Advillin^{CreERT2}; Shank3^{fl/+}* or *Advillin^{CreERT2}; Mecp2^{fl/y}* mutant mice and their control littermates. TAM was administered from P28-32 to all littermates.

(G) Quantification of the number of PV⁺ neurons in S1 of *Advillin^{CreERT2}; Shank3^{fl/+}* or *Advillin^{CreERT2}; Mecp2^{fl/y}* mutant mice and their control littermates. Mice received five days of TAM treatment, beginning at either P5, P10 or P28. Student's t test, * $p < 0.05$.

(H) Quantification of the number of PV⁺ neurons in V1 of *Advillin^{CreERT2}; Shank3^{fl/+}* or *Advillin^{CreERT2}; Mecp2^{fl/y}* mutant mice and their control littermates. Mice received five days of TAM treatment, beginning at either P5, P10 or P28. Student's t test, not significant.

(I) Quantification of the number of PV⁺ neurons in BLA of *Advillin^{CreERT2}; Shank3^{fl/+}* or *Advillin^{CreERT2}; Mecp2^{fl/y}* mutant mice and their control littermates. Mice received five days of TAM treatment, beginning at either P5, P10 or P28. Student's t test, * $p < 0.05$.

(J) IHC images of coronal primary somatosensory cortex (S1), spinal cord, or DRG sections from control and *Advillin^{CreERT2}; Mecp2^{STOP/y}* mice showing MeCP2, Hoechst and IB4 expression with and without the TAM administration ('+ TAM', '- TAM'). TAM was administered from P28-32.

(K) Quantification of the number of PV⁺ neurons in S1 of control, mutant and mutant rescue mice. Mice received five days of TAM treatment, beginning at P28. One-way ANOVA with post hoc Tukey's test, [for *Shank3* groups: $F(2,9) = 6.206$, $p = 0.0202$; for *Mecp2* groups: $F(2,10) = 30.95$, $p < 0.0001$], * $p < 0.05$.

(L) Quantification of the number of PV⁺ neurons in V1 of control, mutant and mutant rescue mice. Mice received five days of TAM treatment, beginning at P28. One-way ANOVA with post hoc Tukey's test, [for *Shank3* groups: $F(2,9) = 5.794$, $p = 0.0241$; for *Mecp2* groups: $F(2,10) = 30.45$, $p < 0.0001$], * $p < 0.05$.

(M) Quantification of the number of PV⁺ neurons in BLA of control, mutant and mutant rescue mice. Mice received five days of TAM treatment, beginning at P28. One-way ANOVA with post hoc Tukey's test, [for *Shank3* groups: $F(2,9) = 25.88$, $p = 0.0002$; for *Mecp2* groups: $F(2,10) = 13.24$, $p < 0.0015$], * $p < 0.05$. Error bars indicate standard error of the mean.

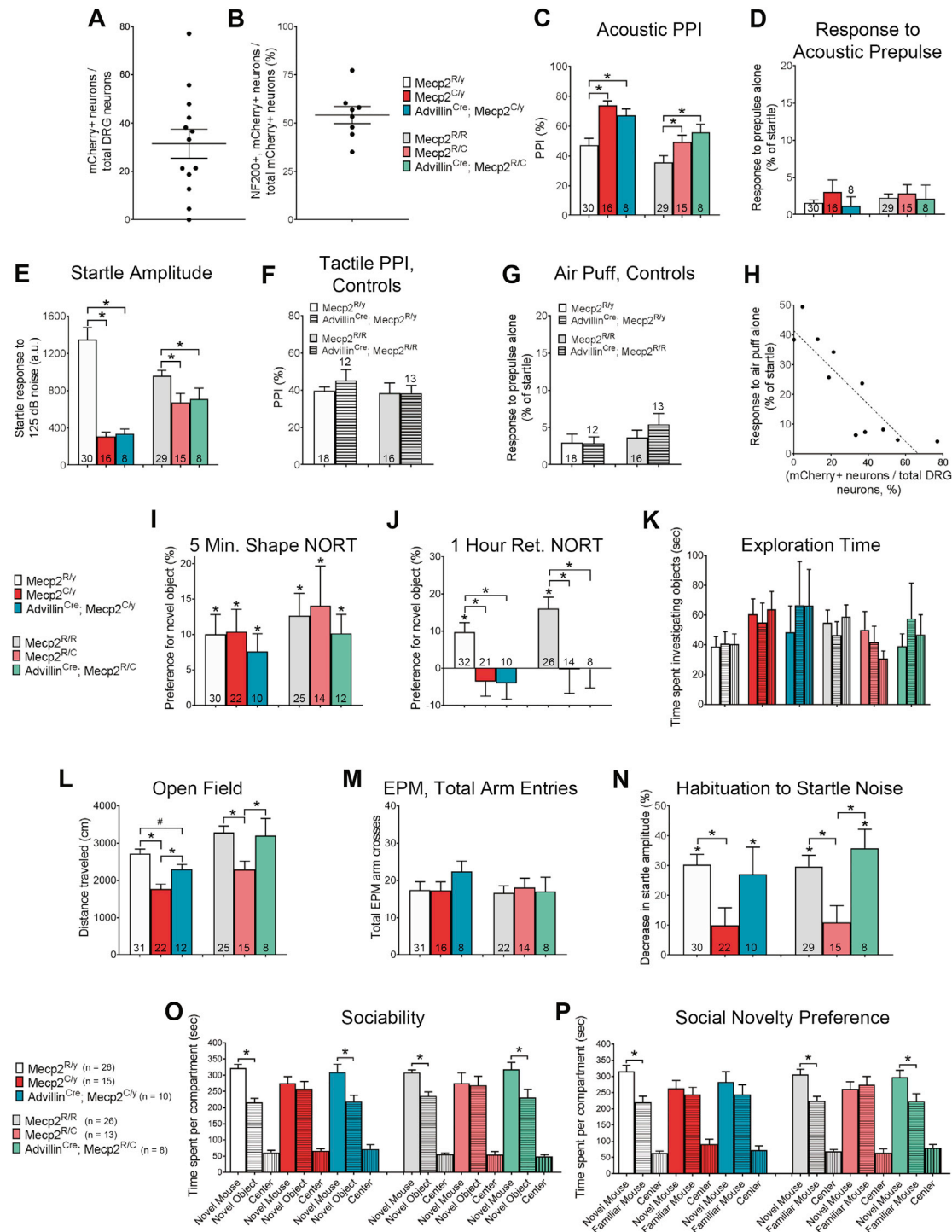
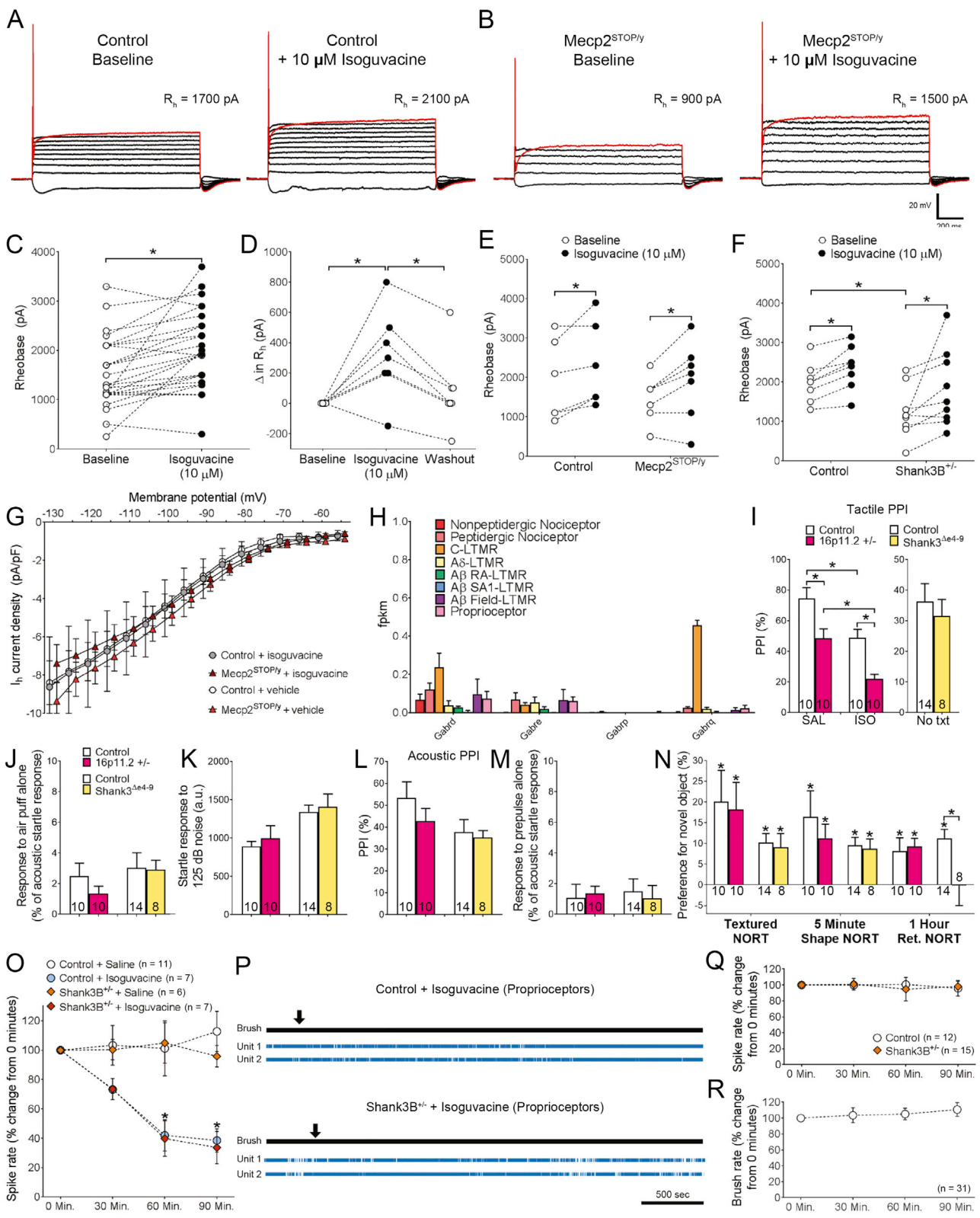


Figure S5. Viral Expression of GABRB3 in Peripheral Sensory Neurons, Beginning at P5, Improves Some Tactile, Brain, and Behavioral Deficits Observed in *Mecp2^{R306C}* Mutant Mice, Related to Figure 4

(A) Percentage of transduced DRG neurons (mCherry⁺ neurons) in mice that received P5 i.p. injection of AAV.FLEX.GABRB3.mCherry
 (B) Percentage of mCherry⁺ DRG neurons that are NF200+ neurons in mice that received P5 i.p. injection of AAV.FLEX.GABRB3.mCherry
 (C) Percent inhibition of the startle response to a 125 dB noise (pulse), when the startle noise is preceded by tone prepulse in male and female control (*Mecp2^{R/ly}* or *R/R*), mutant (*Mecp2^{C/ly}* or *R/C*) and mutant rescue (*Advillin^{Cre}; Mecp2^{C/ly}* or *R/C*) mice. All littermates received an i.p. injection of AAV.FLEX.GABRB3.mCherry at P5. One-way ANOVA with post hoc Tukey's test, **p* < 0.05.

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- (D) Response to a non-startling acoustic noise (80 dB, 20 ms) in male and female control, mutant and mutant rescue mice. Responses are expressed as percent of startle response to a 125 dB startle noise. One-way ANOVA, not significant.
- (E) Magnitude of startle response to a 125 dB noise in male and female control, mutant and mutant rescue mice. One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (F) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in *Mecp2^{R/y}* and *Advillin^{Cre}; Mecp2^{R/y}* control mice. Student's unpaired t test, not significant.
- (G) Response to a light air puff stimulus alone in *Mecp2^{R/y} or R/R* and *Advillin^{Cre}; Mecp2^{R/y} or R/R* control mice. Responses are expressed as percent of startle response to a 125 dB noise. Student's unpaired t test, not significant.
- (H) Percent of DRG neurons transduced with AAV.FLEX.GABRB3.mCHERRY, plotted against response to a light air puff stimulus alone for *Advillin^{Cre}; Mecp2^{C/y}* mice. Linear regression analysis [$F(1,9) = 27.30$, $P = 0.0005$]; $R^2 = 0.7520$.
- (I) Discrimination index for 5 minute shape NORT. For comparisons between groups, one-way ANOVA, not significant.
- (J) Discrimination index for 1 hour retention NORT. One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (K) Average amount of time (seconds) spent physically interacting with both the familiar and novel object in the NOR tests in mutant mice and their control littermates. Bars with no lines: 5 minute shape NORT; bars with horizontal lines: textured NORT; bars with vertical lines: 1 hour retention NORT. One-way ANOVA, not significant.
- (L) Average total distance traveled in the open field chamber for mutant, mutant rescue mice and their control littermates. One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (M) Average number of EPM arm entries during the 10-minute assay, for mutant mice and their control littermates. One-way ANOVA, not significant.
- (N) Percent decrease in startle response to a 125 dB noise during a 30-minute tactile PPI session, when comparing the first five startle responses to the last five responses to a 125 dB noise for mutant, mutant rescue mice and their control littermates. One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (O) Preference index for the percentage of time spent investigating a novel mouse, compared to a novel object, in the "Sociability" portion of the 3-chamber social interaction test. One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (P) Preference index for the percentage of time spent investigating a novel mouse, compared to a familiar mouse, in the "Social Novelty Preference" portion of the 3-chamber social interaction test. One-way ANOVA with post hoc Tukey's test, * $p < 0.05$. Error bars indicate standard error of the mean.



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Figure S6. Isoguvacine Attenuates Tactile Sensitivity through Reduced Excitability of Peripheral, Low-Threshold Mechanosensory Neurons but Does Not Affect Proprioceptor Firing Rates, Related to Figures 5 and 6

(A-B) Representative traces from large diameter neurons cultured from DRGs of control (A) and *Mecp2*^{STOP/y} (B) mice during whole cell current clamp recordings, in which the minimal amount of current required to elicit an action potential in each neuron (rheobase, R_h), was determined in the absence or presence of 10 μ M isoguvacine.

(C) Quantification of average R_h in large diameter neurons cultured from DRGs of control mice during a baseline period and then in presence of 10 μ M isoguvacine. Paired t test, * $p < 0.0005$.

(D) Quantification of average R_h in large diameter neurons cultured from DRGs of control mice during a baseline period, followed application of 10 μ M isoguvacine, then washout period. Repeated-measures one-way ANOVA with post hoc Sidak's test [F (5,10) = 20.63, $p < 0.0001$], * $p < 0.05$.

(E) Quantification of average R_h in large diameter neurons cultured from DRGs of control and *Mecp2*^{STOP/y} mice during a baseline period and then in presence of 10 μ M isoguvacine. Repeated-measures two-way ANOVA with post hoc Sidak's test [F (11,11) = 20.63, $p < 0.0001$], * $p < 0.05$.

(F) Quantification of average R_h in large diameter neurons cultured from DRGs of control and *Shank3B*^{+/-} mice during a baseline period and then in presence of 10 μ M isoguvacine. Repeated-measures two-way ANOVA with post hoc Sidak's test [F (15,15) = 4.545, $p = 0.0029$], * $p < 0.05$.

(G) Quantification of total I_h density at each hyperpolarizing voltage step for large diameter neurons cultured from DRGs of *Mecp2*^{STOP/y} mutant mice and control littermates, in the presence of vehicle or 10 μ M isoguvacine. Repeated-measures two-way ANOVA [F (3, 1280) = 1.953, $p = 0.1193$], not significant.

(H) GABA_A receptor subunit RNA expression levels across peripheral somatosensory neuron subtypes, for the delta, epsilon, pi, and theta subunit types.

(I) Percent inhibition of the startle response to a 125 dB noise, when the startle noise is preceded by a light air puff in 16p11.2 deletion mutant mice (16p11.2^{+/-}) or heterozygous *Shank3* ^{Δ ex4-9} null mice and their control littermates. 16p11.2^{+/-} mice received i.p. administration of either saline ('SAL') or 2 mg/kg isoguvacine ('ISO'). *Shank3* ^{Δ ex4-9} mice received no treatment ('No txt'). For 16p11.2^{+/-} mice: repeated-measures, two-way ANOVA with post hoc Tukey's test [F (2,36) = 14.86, $p < 0.0001$], * $p < 0.05$. For *Shank3* ^{Δ ex4-9} mice: Student's unpaired t test, not significant.

(J) Response to a light air puff stimulus alone in 16p11.2^{+/-} or heterozygous *Shank3* ^{Δ ex4-9} mutant mice and their control littermates. Responses are expressed as percent of startle response to a 125 dB noise in 16p11.2^{+/-} or heterozygous *Shank3* ^{Δ ex4-9} mutant mice and their control littermates. Student's unpaired t test, not significant.

(K) Magnitude of startle response to a 125 dB noise in 16p11.2^{+/-} or heterozygous *Shank3* ^{Δ ex4-9} mutant mice and their control littermates. Student's unpaired t test, not significant.

(L) Percent inhibition of the startle response to a 125 dB noise (pulse), when the startle noise is preceded by tone prepulse (80 dB, 'acoustic PPI') in 16p11.2^{+/-} or heterozygous *Shank3* ^{Δ ex4-9} mutant mice and their control littermates. Student's unpaired t test, not significant.

(M) Response to a non-startling acoustic noise (80 dB, 20 ms), in 16p11.2^{+/-} or heterozygous *Shank3* ^{Δ ex4-9} mutant mice and their control littermates. Responses are expressed as percent of startle response to a 125 dB startle noise. Student's unpaired t test, not significant.

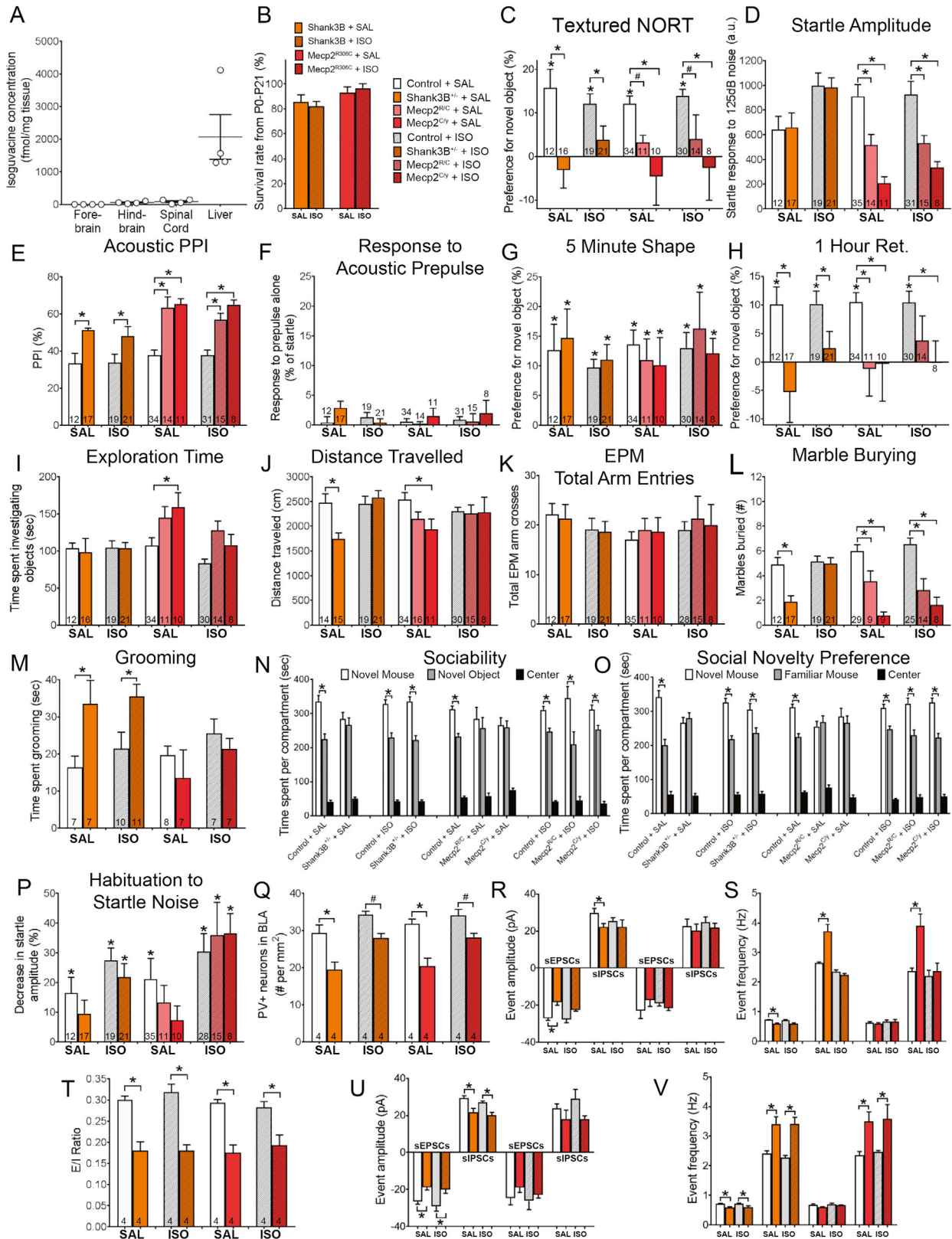
(N) Discrimination index for textured, 5 minute shape and 1 hour retention NORT in 16p11.2^{+/-} or heterozygous *Shank3* ^{Δ ex4-9} mutant mice and their control littermates. For comparisons between genotypes: Student's unpaired t test, * $p < 0.05$.

(O) Average firing frequency of LTMRs in response to an air puff stimulus over the duration of each recording experiment, following subcutaneous injection of either saline or isoguvacine (2 mg/kg). Repeated-measures, two-way ANOVA with post hoc Dunnett's test [F (3, 108) = 16.02, $p < 0.0001$], * $p < 0.05$.

(P) Representative activity raster plots for multiple putative proprioceptors in multiple mice over the duration of a recording experiment in controls and *Shank3B*^{+/-} mice. Mice received a subcutaneous injection of isoguvacine (2 mg/kg) during the experiment, and activity of proprioceptive units in response to a brush stimulus was assessed over a 90-minute period. Arrows indicate time of injection.

(Q) Average firing frequency of proprioceptors in response to a brush stimulus over the duration of each recording experiment in controls and *Shank3B*^{+/-} mice, following subcutaneous injection isoguvacine (2 mg/kg). Repeated-measures, two-way ANOVA with post hoc Dunnett's test [F (3, 100) = 0.01423, $p = 0.9053$], not significant.

(R) Average brush stroke frequency over the duration of each recording experiment in all controls and mutant mice, following subcutaneous injection of either saline or isoguvacine (2 mg/kg). Repeated-measures, one-way ANOVA with post hoc Dunnett's test [F (3,44) = 0.3736, $p = 0.7725$], not significant. Error bars indicate standard error of the mean.



(legend on next page)

Figure S7. Chronic administration of Isoguvacine Improves Tactile Over-Reactivity, Region-Specific Cortical Abnormalities, and Some ASD-Related Behaviors in *Mecp2* and *Shank3* Mutant Mice, Related to Figure 7

- (A) Liquid chromatography mass spectrometry (LC-MS) quantified isoguvacine concentrations in brain homogenate (forebrain and hindbrain), spinal cord and liver samples of mice treated with isoguvacine (2 mg/kg, i.p.) from P1-42. Samples were obtained six hours following the final day of isoguvacine administration.
- (B) Percent of pups surviving to P21 in litters from *Shank3B^{+/-}* or *Mecp2^{R/C}* female mice, in which litters were treated daily with either saline or isoguvacine (2 mg/kg). Student's unpaired t test, not significant.
- (C) Discrimination index for textured NORT. One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (D) Magnitude of startle response to a 125 dB noise in *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (E) Percent inhibition of the startle response to a 125 dB noise (pulse), when the startle noise is preceded by tone prepulse in *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (F) Response to a non-startling acoustic noise (80 dB, 20 ms) in *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). Responses are expressed as percent of startle response to a 125 dB startle noise. One-way ANOVA with post hoc Tukey's test, not significant.
- (G) Discrimination index for 5 minute shape NORT in *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). For comparisons between groups: one-way ANOVA with post hoc Tukey's test, not significant.
- (H) Discrimination index for 1 hour retention NORT in *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (I) Average amount of time (seconds) spent physically interacting with both the familiar and novel object in the NOR tests *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (J) Average total distance traveled in the open field chamber for *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (K) Average number of EPM arm entries during the 10-minute assay, for *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA, not significant.
- (L) Average number of marbles buried (out of 12) during a twenty-minute assay for *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (M) Average amount of time spent grooming during a 10-minute open field assessment for *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (N) Time spent per compartment in the 3-chamber social interaction test during the "Sociability" portion of the assay for *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (O) Time spent per compartment in the 3-chamber social interaction test during the "Social Novelty Preference" portion of the assay for *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (P) Percent decrease in startle response to a 125 dB noise during a 30-minute tactile PPI session, when comparing the first five startle responses to the last five responses to a 125 dB noise for *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). For comparisons between groups: one-way ANOVA, not significant.
- (Q) Quantification of the number of PV⁺ neurons in BLA of *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$, # $p < 0.10$.
- (R) Quantification of sEPSC and sIPSC event amplitude in S1 slices from *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). Two-way ANOVA with post hoc Sidak's test, * $p < 0.05$.
- (S) Quantification of sEPSC and sIPSC event frequency in S1 slices from *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). Two-way ANOVA with post hoc Sidak's test, * $p < 0.05$.
- (T) Quantification of excitatory/inhibitory (E/I) ratios in V1 slices from *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). One-way ANOVA with post hoc Tukey's test, * $p < 0.05$.
- (U) Quantification of sEPSC and sIPSC event amplitude in V1 slices from *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). Two-way ANOVA with post hoc Sidak's test, * $p < 0.05$.
- (V) Quantification of sEPSC and sIPSC event frequency in V1 slices from *Shank3B^{+/-}*, *Mecp2^{R/C}* or *Mecp2^{C/y}* mutant mice and control littermates treated daily from P1-42 with either saline or isoguvacine (2 mg/kg). Two-way ANOVA with post hoc Sidak's test, * $p < 0.05$. Error bars indicate standard error of the mean.